

<https://helda.helsinki.fi>

On Sistotremastrum and similar-looking taxa (Trechisporales, Basidiomycota)

Spirin, Viacheslav

2021-04

Spirin , V , Volobuev , S , Viner , I , Miettinen , O , Vlasák , J , Schoutteten , N ,
Motato-Vásquez , V , Kotiranta , H , Hernawati , N & Larsson , K-H 2021 , ' On
Sistotremastrum and similar-looking taxa (Trechisporales, Basidiomycota) ' , Mycological
Progress , vol. 20 , no. 4 , pp. 453-476 . <https://doi.org/10.1007/s11557-021-01682-z>

<http://hdl.handle.net/10138/330628>

<https://doi.org/10.1007/s11557-021-01682-z>

CC BY

publishedVersion

Downloaded from Helda, University of Helsinki institutional repository.

This is an electronic reprint of the original article.

This reprint may differ from the original in pagination and typographic detail.

Please cite the original version.



On *Sistotremastrum* and similar-looking taxa (Trechisporales, Basidiomycota)

Viacheslav Spirin¹ · Sergey Volobuev² · Ilya Viner¹ · Otto Miettinen¹ · Josef Vlasák³ · Nathan Schoutteten⁴ · Viviana Motato-Vásquez⁵ · Heikki Kotiranta⁶ · Hernawati^{7,8} · Karl-Henrik Larsson^{9,10}

Received: 20 October 2020 / Revised: 28 December 2020 / Accepted: 2 February 2021
© The Author(s) 2021

Abstract

The taxonomy of *Sistotremastrum* (Trechisporales, Basidiomycota) is revised based on morphology and DNA data. The genus is shown to be polyphyletic, and therefore it is split into two units—*Sistotremastrum* s. str. and *Sertulicium*, gen. nov. (typified with *Corticium niveocreum*). *Sistotremastrum* s. str. is retained for eleven species of which eight are described as new while *Sertulicium* encompasses at least six species, including one new to science. Both of these genera are only distantly related to other representatives of the Trechisporales. Additionally, a new poroid neotropical species, *Porpomyces abiens* (Hydnodontaceae), is described as morphologically similar to some members of *Sistotremastrum* s. str.

Keywords Corticioid fungi · Phylogeny · Taxonomy · New taxa

Introduction

Sistotremastrum J. Erikss. is a genus of corticioid fungi typified with *S. suecicum* Litsch. ex J. Erikss. (Eriksson 1958). Initially introduced for two species, it has been gradually expanded to encompass smooth, effused basidiomycetes with clavate basidia producing four to six sterigmata and smooth, thin-walled, inamyloid basidiospores (Boidin and Gilles 1994; Telleria et al. 2013a, 2014). Recently, Gruhn et al. (2018) described two new *Sistotremastrum* species with a hydroid hymenophore and constantly four-sterigmatic basidia

and thus changed the present concept of the genus. As a consequence, the morphological delimitation of *Sistotremastrum* from *Brevicellicium* K.H. Larsson & Hjortstam and, partly, from *Trechispora* P. Karst. became obscure. Another yet persisting problem is the separation of *Sistotremastrum* from *Paullicorticium* J. Erikss., which has a similar set of microscopic characters (Eriksson et al. 1984). Oberwinkler (1965) moved *Sistotremastrum niveocreum* (Höhn. & Litsch.) J. Erikss., one of the original species, to *Paullicorticium*, stressing similarities in basidial development. Larsson et al. (2004) included *Paullicorticium ansatum* Libert in

Section Editor: Yu-Cheng Dai

✉ Viacheslav Spirin
viacheslav.spirin@helsinki.fi

¹ Finnish Museum of Natural History, University of Helsinki, PO Box 7, 00014 University of Helsinki, Helsinki, Finland

² Komarov Botanical Institute RAS, 2 Prof. Popov str, St. Petersburg 197376, Russia

³ Biology Centre, Academy of Sciences of the Czech Republic, Branišovská 31, 37005 České Budějovice, CZ, Czech Republic

⁴ Research Group Mycology, Department of Biology, Ghent University, K.L. Ledeganckstraat 35, B-9000 Gent, Belgium

⁵ Núcleo de Pesquisa em Micologia, Instituto de Botânica, Av. Miguel Estéfano 3687, São Paulo, SP 04301-902, Brazil

⁶ Finnish Environment Institute, Latokartanonkaari 11, 00790 Helsinki, Finland

⁷ The Herbarium (ANDA), Universitas Andalas, Padang, Sumatera Barat 25163, Indonesia

⁸ Fakultas Kehutanan (Faculty of Forestry), Universitas Muhammadiyah Sumatera Barat, Padang, Sumatera Barat 25172, Indonesia

⁹ Natural History Museum, University of Oslo, PO Box 1172, Blindern, 0318 Oslo, Norway

¹⁰ Gothenburg Global Biodiversity Centre, Post Box 461, 40530 Gothenburg, Sweden

phylogenetic analyses of Agaricomycetes but were unable to place it in one of the clades currently recognized as orders. On the other hand, all *Sistotremastrum* species, including the generic type, have been found to be members of the Trechisporales (Larsson 2007; Telleria et al. 2013a, 2014; Gruhn et al. 2018). However, most *Paullicorticium* species have not yet been studied by DNA methods—in particular, *P. jacksonii* Liberta whose connection to *Sistotremastrum* was discussed in the older literature (Eriksson et al. 1978). In the present paper, we revise the generic limits of *Sistotremastrum* and similar-looking taxa, and investigate species diversity based on collections from different regions of Eurasia, as well as from North and South America.

Materials and methods

Morphological study

Type specimens and collections from herbaria H, O, LE, S, GB, MA, W, GENT, TRTC, LY, CWU, ANDA, MAN, SP, MG, SING, as well as from the private herbarium of the author JV were studied. Herbarium acronyms are given according to Thiers (2019). Microscopic methods follow Miettinen et al. (2018). All measurements were made from microscopic slides mounted in Cotton Blue (abbreviated as CB in descriptions below), using phase contrast and oil immersion lens (Leitz Diaplan microscope, × 1250 amplification). In total, 20–30 basidiospores, 20 basidia and subhymenial/subicular hyphae, and at least 10 basidioles and hyphidia were measured for each specimen studied. The following abbreviations are used in morphological descriptions: L—mean basidiospore length, W—mean basidiospore width, Q'—length/width ratio, Q—mean length/width ratio, n—number of measurements per specimens.

DNA extraction, amplification, and sequencing

DNA was extracted from herbarium specimens using the EZNA Forensic DNA kit (Omega Bio-tek) and AxyPrep Multisource Genomic DNA Miniprep kit (Axygen Biosciences, CA, USA) according to the manufacturer's instructions, except that 50 µl of the elution buffer was used in the elution step of the last one. The ribosomal ITS1–5.8S–ITS2 region was amplified and sequenced with the fungal specific primers ITS1F and ITS4B (Gardes and Bruns 1993). Sequences of nrLSU-rDNA were generated using primers LR0R, CTB6, LR5, and LR7 (Vilgalys and Hester 1990; Haynes et al. 1995). PCR products were visualized using agarose gel electrophoresis and Gel Red staining, and subsequently purified with the Fermentas Genomic DNA Purification Kit (Thermo Fisher Scientific, MA, USA). The resulting products were sequenced with an ABI model 3130 genetic analyzer (Applied Biosystems, CA, USA) and

BigDye v.3.1 and ABI3730XL analyzer (Applied Biosystems) by Macrogen. The raw data were edited and assembled in MEGA 6 (Tamura et al. 2013).

Phylogenetic analyses

For this study, we generated 60 nrITS and 36 nrLSU sequences. All other sequences used in the analyses were downloaded from GenBank (Benson et al. 2018) or UNITE (Nilsson et al. 2018) (Table 1). All newly generated sequences were deposited in the INSDC.

Extremely high diversity of ITS sequences in the studied taxa precludes attempts to construct a reliable all-encompassing alignment for all *Sistotremastrum* spp. However, we could distinguish five alignable groups: *S. niveocremaum* complex, *S. suecicum* and its closest relatives, *S. aculeocremitans* and its closest relatives, *S. fibrillosum* complex, and *S. rigidum*–*vigilans* clade. We found it reasonable to exclude *S. fibrillosum*, as well as newly described *S. rigidum* and *S. vigilans* from ITS analysis since molecular evidence provided by nrLSU analysis is sufficient for the purpose of our study. Therefore, we generated four alignments for this study: (1) nrLSU-alignment for the Trechisporales, (2) nrITS-alignment for *S. niveocremaum*, (3) nrITS-alignment limited to *S. suecicum* and its closest relatives, and (2) nrITS-alignment for *S. aculeocremitans* and its closest relatives. The alignments were calculated through MAFFT 7.429 online server (<https://mafft.cbrc.jp/alignment/server/>) using the L-INS-I strategy (Katoh et al. 2017) and then manually adjusted. The alignments are deposited in TreeBASE (S25768).

For the genus-level analysis, a nrLSU dataset (1) was assembled with representatives of Trechisporales (65 sequences). The resulting tree was rooted with *Onnia leporina* and *Sphagnomphalia brevibasidiata* (Hymenochaetales) and *Sphaerobolus stellatus* (Geastrales, Phallomycetidae). This choice was guided by the current JGI Basidiomycota tree (https://mycocosm.jgi.doe.gov/mycocosm/species-tree/tree;_FJDxL?organism=basidiomycota) where Trechisporales is recovered close to Hymenochaetales and Phallomycetidae. After removing unalignable, ambiguous alignment positions, the alignment length was 742 bp with 184 variable site patterns. The three ITS datasets were assembled similarly: the *S. niveocremaum* species complex alignment (2) contained 60 sequences and had the length of 329 bp with 40 variable site patterns after removing unalignable, ambiguous positions. The *S. suecicum* species complex analysis (3) contained 22 sequences, with alignment length of 508 bp with 13 variable site patterns after removing ambiguous positions. The *S. aculeocremitans* species complex analysis (4) contained 22 sequences with the alignment length of 470 bp with 37 variable site patterns. All the ITS trees are midpoint-rooted.

We inferred phylogenetic trees with maximum likelihood (ML), maximum parsimony (MP), and Bayesian inference (BI) but show here only the latter since all trees show

Table 1 Sequences generated for the present study

Species	Specimen/herbarium	Geographic origin (ISO code)	Host	GenBank number	
				nrITS	nrLSU
<i>Brevicellicium exile</i>	Spirin 8370 (H)	US-WA	<i>Tsuga heterophylla</i>	MT002322	MT002338
<i>B. olivascens</i>	Spirin 4446 (H)	RU-NIZ	<i>Quercus robur</i>	MT002327	-
<i>B. viridulum</i>	Kotiranta 29271 (H)	RU-SAK	<i>Alnus hirsuta</i>	MN983275	-
<i>Luellia cystidiata</i>	Læssøe 13875 (GB)	DK	<i>Picea abies</i>	MW371211	MW371211
<i>Porpomyces abiens</i>	Vlasák 1808/16 (H)	GF	Hardwood	MN987945	MN987945
<i>P. abiens</i>	Vlasák 1808/39 (H)	GF	Hardwood	MN987942	MN987942
<i>Pteridomyces galzinii</i>	Bernicchia 8122 (GB)	IT	<i>Polystichum aculeatum</i>	MN937559	MN937559
<i>Sertulicium granuliferum</i>	Spirin 9296 (H)	RU-NIZ	<i>Populus tremula</i>	MT002321	-
<i>S. granuliferum</i>	LE 292194	RU-ORL	<i>Betula pendula</i>	MT002325	MT002333
<i>S. granuliferum</i>	LE 314053	RU-ORL	<i>Q. robur</i>	MT002313	MT002340
<i>S. granuliferum</i>	LE 314054	RU-ORL	<i>Q. robur</i>	MT002315	MT002342
<i>S. granuliferum</i>	LE 314055	RU-ORL	<i>B. pendula</i>	MT002314	MT002341
<i>S. granuliferum</i>	LE 314056	RU-ORL	<i>Q. robur</i>	MT002316	MT002343
<i>S. granuliferum</i>	LE 314057	RU-ORL	<i>Q. robur</i>	MT002312	MT002339
<i>S. granuliferum</i>	Kotiranta 26776 (H)	RU-TY	<i>Larix sibirica</i>	MT002323	-
<i>S. granuliferum</i>	CWU 4704	UA	<i>Salix alba</i>	MT002326	-
<i>S. granuliferum</i>	Miettinen 14813.2 (H)	US-MA	Cut bole	MT075854	-
<i>S. granuliferum</i>	Miettinen 16083 (H)	US-MA	Hardwood	MT002319	-
<i>S. granuliferum</i>	Miettinen 17272 (H)	US-MA	<i>Quercus</i> sp.	MT002330	-
<i>S. granuliferum</i>	Larsson 12282 (GB)	US-TN	Decayed wood	MN937560	MN937560
<i>S. granuliferum</i>	Larsson 12297 (GB)	US-TN	Decayed wood	MN937561	MN937561
<i>S. jacksonii</i>	Svantesson 699 (O)	NO	<i>Picea abies</i>	MN937562	MN937562
<i>S. jacksonii</i>	Spirin 10425 (H)	RU-LEN	<i>P. abies</i>	MN987943	MN987943
<i>S. jacksonii</i>	Miettinen 17141 (H)	US-NY	<i>Abies</i> sp.	MN987944	-
<i>S. lateclavigerum</i>	Spirin 13457	SI	<i>P. abies</i>	MW049161	-
<i>S. niveocreum</i>	Van Autgaerden S-20 (GENT)	BE	Hardwood	MN947227	MN930920
<i>S. niveocreum</i>	Miettinen 14925.3 (H)	FI	<i>Sorbus aucuparia</i> (?)	MT075855	-
<i>S. niveocreum</i>	Söderholm 4050 (H)	FI	<i>Salix</i> sp.	MT075856	MT002334
<i>S. niveocreum</i>	Larsson 13727 (GB)	FR	Hardwood	MN937563	MN937563
<i>S. niveocreum</i>	Kotiranta 26267 (H)	RU-KRA	<i>Alnus sibirica</i>	MT075858	MT002337
<i>S. vernale</i>	Söderholm 3886 (H)	FI	<i>P. abies</i>	MT002311	MT664174
<i>Sertulicium</i> sp. 1	Spirin 5158 (H)	RU-KHA	<i>Corylus mandshurica</i>	MT075857	MT002344
<i>Sistotremastrum aculeatum</i>	Miettinen 10380.1 (H)	CN-YN	Hardwood (?)	MN991176	MW045423
<i>S. aculeatum</i>	Miettinen 13799.1 (ANDA)	ID-SB	Hardwood	MN988623	-
<i>S. aculeatum</i>	Dunaev KUN 1105 (H)	TH	Hardwood	MN991181	-
<i>S. aculeicrepitans</i>	Larsson 16097 (URM)	BR-PB	Hardwood	MN937564	MN937564
<i>S. aculeicrepitans</i>	Larsson 16478 (MG)	BR-PA	Hardwood	MN937565	-
<i>S. confusum</i>	Larsson 16004 (URM)	BR-PE	Hardwood	MN937567	MN937567
<i>S. confusum</i>	Larsson 16023 (URM)	BR-PE	Hardwood	MN937566	-
<i>S. denticulatum</i>	Motato-Vásquez 894 (SP)	BR-SP	Fallen branch	MN954694	MW045424
<i>S. fibrillosum</i>	Larsson 16988 (MG)	BR-PA	Hardwood	MN937568	MN937568
<i>S. geminum</i>	Miettinen 14333 (MAN)	ID-PB	<i>Intsia bijuga</i>	MN991177	MN991177
<i>S. induratum</i>	Spirin 8598 (H)	US-WA	<i>Abies grandis</i>	MT002324	MT664173
<i>S. induratum</i>	Spirin 8804 (H)	US-WA	<i>Picea sitchensis</i>	MT002318	-
<i>S. mendax</i>	J. Nordén 9579 (O)	NO	<i>P. abies</i>	MN937569	-
<i>S. mendax</i>	Larsson 12022 (O)	NO	<i>P. abies</i>	MN937570	MN937570
<i>S. mendax</i>	Miettinen 20946 (H)	RU-LEN	<i>P. abies</i>	MN991179	-

Table 1 (continued)

Species	Specimen/herbarium	Geographic origin (ISO code)	Host	GenBank number	
				nrITS	nrLSU
<i>S. rigidum</i>	Motato-Vásquez 833 (SP)	BR-SP	Fallen log	MN954693	MW045435
<i>S. suecicum</i>	Spirin 8932 (H)	CA-BC	<i>Pinus contorta</i>	MT002317	-
<i>S. suecicum</i>	Kunttu 5959 (H)	FI	<i>Pinus sylvestris</i>	MT075859	MT002335
<i>S. suecicum</i>	LE 295792	RU-ARK	<i>P. sylvestris</i>	MT002332	-
<i>S. suecicum</i>	Miettinen 14550.1 (H)	SE	<i>P. sylvestris</i>	MT075860	MT002336
<i>S. suecicum</i>	Larsson 11849 (GB)	SE	<i>P. abies</i> (?)	MN937571	MN937571
<i>S. suecicum</i>	Miettinen 14829 (H)	US-MA	Fallen branch	MT075861	-
<i>S. suecicum</i>	Miettinen 16061 (H)	US-MA	<i>Tsuga canadensis</i>	MT002328	-
<i>S. suecicum</i>	Miettinen 16618 (H)	US-MA	<i>Tsuga</i> sp./ <i>Pinus</i> sp.	MT002331	-
<i>S. vigilans</i>	Fonneland 2011-78 (O)	NO	<i>P. abies</i>	MN937572	MN937572
<i>S. vigilans</i>	Spirin 10097 (H)	RU-LEN	<i>P. abies</i>	MN991178	-
<i>S. vigilans</i>	Spirin 8778 (H)	US-WA	<i>T. heterophylla</i>	MN991182	MN991182
<i>Suillosporium cystidiatum</i>	Spirin 3830 (H)	RU-KHA	<i>Picea ajanensis</i>	MN937573	MN937573

congruity of the phylogenetic signal. The best nucleotide substitution model was chosen with TOPALi 2.5 (Milne et al. 2008) based on the Akaike information criterion (AIC), which were GTR + G + I (nst = 6, rates = invgamma) for the nrLSU-dataset, GTR+G (nst = 6, rates = gamma) for the *S. niveocreum* dataset, and F81 (nst = 1) for the *S. suecicum* dataset. The suggested models were implemented in the Bayesian phylogenetic analyses. We performed Bayesian inference with MrBayes 3.2 (Ronquist et al. 2012). In the analyses, three parallel runs with four chains each, temp = 0.2, were run for 3 million generations. All chains converged to <0.01 average standard deviation of split frequencies. A burn-in of 25% was used in the final analyses. Maximum likelihood (ML) analysis was performed in RAxML 7.2.8 (Stamatakis 2006) implemented in Geneious version 9.1.8 (<http://www.geneious.com>) (Kearse et al. 2012). Following models suggested by TOPALi 2.5, we preferred to use the GTR model with gamma correction (GTR GAMMA) in ML analysis. Bootstrapping was performed using the “Rapid bootstrapping” algorithm with the number of bootstrap replicates set as 1000. Maximum parsimony (MP) analysis was performed using SeaView version 4 (Gouy et al. 2010) with the ignoring all gap sites option. The number of bootstrap replicates was set as 1000.

Results

The nrLSU dataset encompasses all known genera of the Trechisporales as defined by Larsson et al. (2004) and Larsson (2007), with addition of *Pteridomyces* and *Suillosporium*. The overall topologies of the Trechisporales

were highly similar (Fig. 1). They split the order into three strongly supported clades:

- (A) the *Sistotremastrum* s. str. subclade (pp = 1, bs = 99%) includes the generic type, *S. suecicum*, and two unnamed closely related species (introduced below as *S. induratum* and *S. mendax*), all with six-sterigmatic basidia, plus eight more distantly related species with four-sterigmatic basidia (*S. aculeocrepitans*, *S. fibrillosum* and six new species);
- (B) the *S. niveocreum* subclade (pp = 1, bs = 100%) covers five species formerly considered as members of *Sistotremastrum* (except *P. jacksonii* and one unnamed taxon) but separated below into a new genus, *Sertulicium*. Morphological arguments for this solution are given in the taxonomic part of the present paper. *Sistotremastrum guttuliferum* is a younger synonym of *Trechispora granulifera*, and therefore designated as *Sertulicium granuliferum* in the phylogenetic trees; and
- (C) the *Hydnodontaceae* clade (pp = 1, bs = 99%) covers ten genera currently recognized in the family (Larsson 2007; Liu et al. 2019). Among them, two sequences belong to an unnamed poroid fungus from French Guiana introduced below as *Porpomyces abiens*. Anatomically, its basidiocarps are confusingly similar to the four-sterigmatic *Sistotremastrum* species (i.e., *S. aculeocrepitans*, *S. fibrillosum*, and a number of new species described in this study) and differ from them mainly by a truly poroid hymenophore and smaller basidiospores. Sequences of three *Brevicellicium* species (including the generic type, *B. exile*) showing considerable morphological similarity to the four-sterigmatic *Sistotremastrum* species are also resolved in the *Hydnodontaceae*, in accordance to earlier

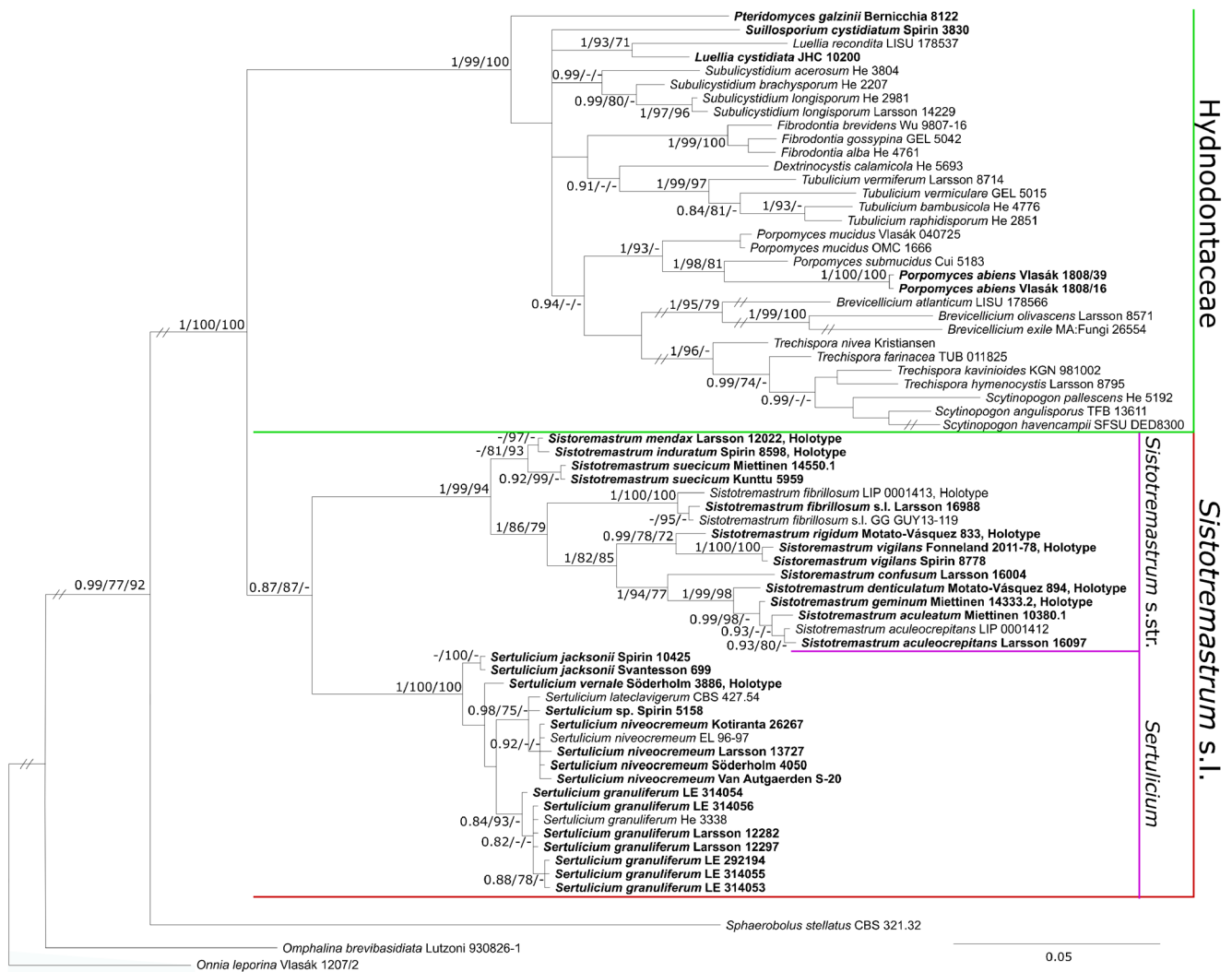


Fig. 1 Phylogenetic relationships of the Trechisporales inferred from nrLSU sequences using Bayesian analysis. A 50% majority rule consensus phylogram. Bayesian posterior probabilities, ML bootstrap,

and MP bootstrap values are shown on nodes; branch lengths reflect estimated number of changes per site

studies (Larsson 2007; Telleria et al. 2013b). Morphological traits differentiating *Sistotremastrum* s. str. from *Brevicellicium* and *Trechispora* are discussed in the taxonomic part of this paper. As for the family-level rearrangement of *Sistotremastrum* and *Sertulicium*, we address this question to further studies with the use of additional genetic markers.

Additionally, three ITS datasets were constructed for the new genus *Sertulicium* and the *Sistotremastrum suecicum* and *S. aculeocreptans* complexes to clarify species limits with the use of larger amount of ITS sequences. The ITS-based topology of *Sertulicium* (Fig. 2) indicates a presence of up to ten species in the genus of which we have named six. Furthermore, ITS sequences of *S. jacksonii* and *S. vernale* reveal some differences (up to 1.4%) between European and

extra-European material. This may indicate that both species, as they are here delimited, actually contain more than one taxon. However, more samples and genetic markers are needed to solve this problem. The second ITS-based phylogeny is restricted to the *S. suecicum* complex (Fig. 3). It shows the presence of at least two more species (described below as *S. induratum* and *S. mendax*) morphologically similar and phylogenetically close to the generic type of *Sistotremastrum*. The third ITS-based phylogram shows phylogenetic relationships of *S. aculeocreptans* and up to five closely related species. Of them, three are described as new to science, all having four-sterigmatic basidia (*S. aculeatum*, *S. denticulatum*, and *S. geminum*) (Fig. 4). Four remaining *Sistotremastrum* species (*S. confusum*, *S. fibrillosum*, *S. rigidum*, and *S. vigilans*) dealt with in the present paper are represented in the nrLSU phylogeny (Fig. 1). Their ITS sequences are available in GenBank (Table 1). Sequenced specimens are marked by asterisk.

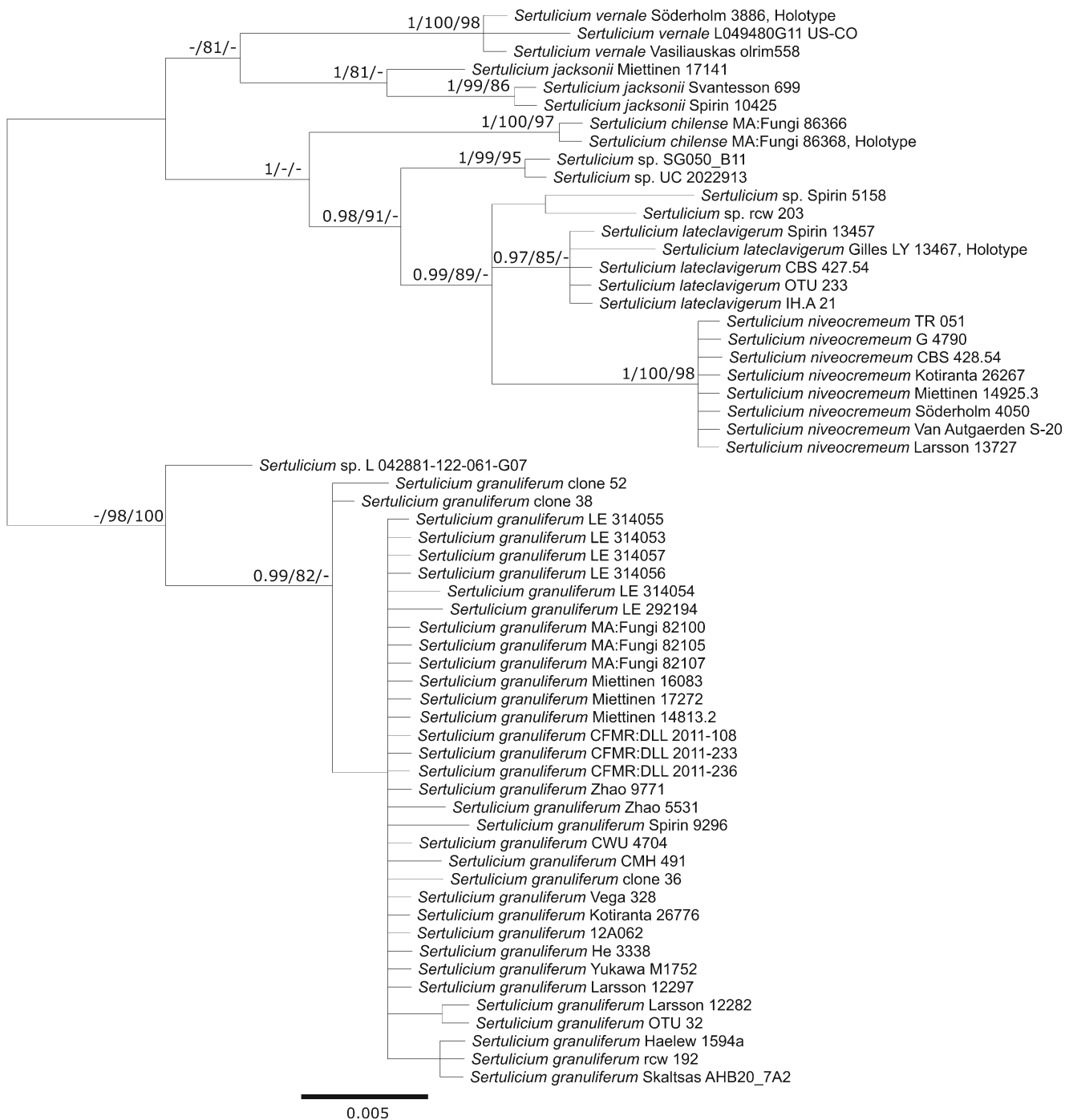


Fig. 2 Phylogenetic relationships of *Sertulicium* inferred from ITS sequences using Bayesian analysis. A 50% majority rule consensus phylogram. Bayesian posterior probabilities, ML bootstrap, and MP

bootstrap values are shown on nodes; branch lengths reflect estimated number of changes per site

Taxonomy

Porpomyces Jülich, Persoonia 11: 425, 1982.

The genus was initially introduced as monotypic, encompassing only the type, *Porpomyces mucidus* (Pers.) Jülich (Jülich 1982). Larsson (2001) showed it is related to *Trechispora*. Another species, *P. submucidus* F. Wu & C.L.

Zhao, was recently described from China (Wu et al. 2015). Here, we introduce the third representative of the genus, *P. abiens* from South America. From *P. mucidus* and *P. submucidus*, *P. abiens* differs by distinctly thick-walled and tightly arranged hyphae in subiculum and trama. This hyphal structure is clearly different from the loosely arranged, rather delicate, thin- or only slightly thick-walled hyphae of

Fig. 3 Phylogenetic relationships of *Sistotremastrum suecicum* and closely related taxa inferred from ITS sequences using Bayesian analysis. A 50% majority rule consensus phylogram. Bayesian posterior probabilities, ML bootstrap, and MP bootstrap values are shown on nodes; branch lengths reflect estimated number of changes per site

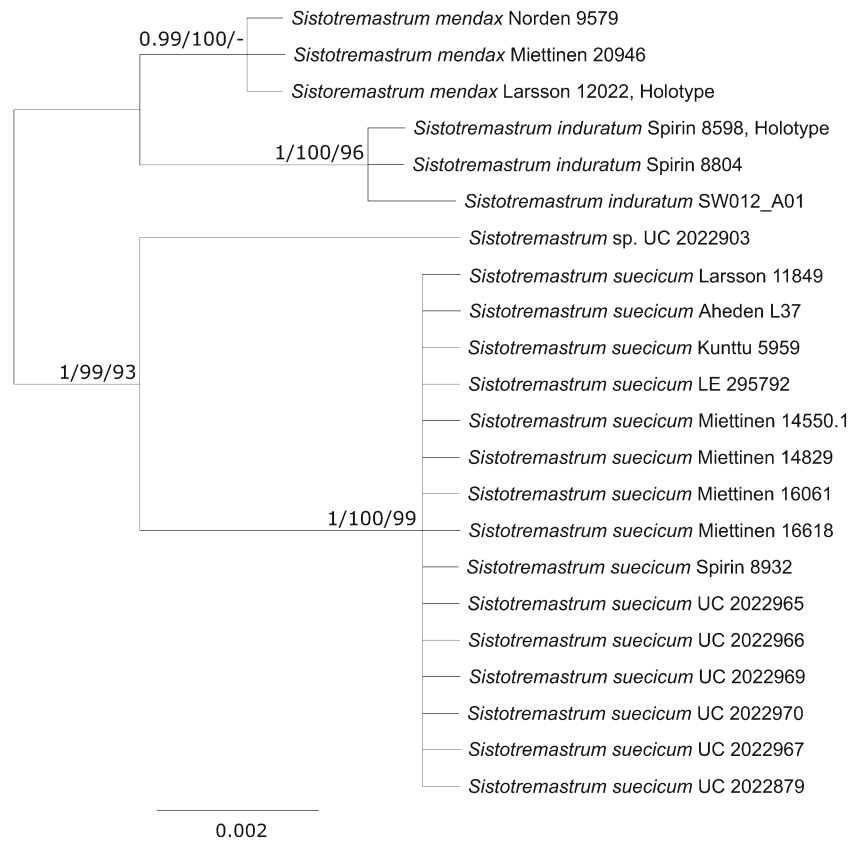
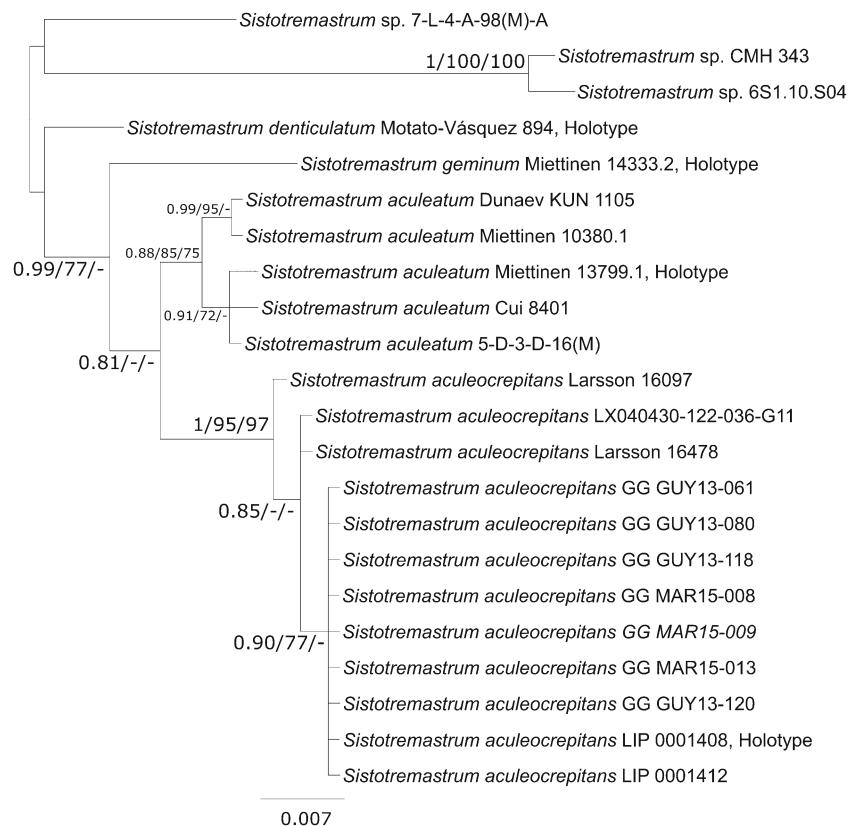


Fig. 4 Phylogenetic relationships of *Sistotremastrum aculeocreptans* and closely related taxa inferred from ITS sequences using Bayesian analysis. A 50% majority rule consensus phylogram. Bayesian posterior probabilities, ML bootstrap, and MP bootstrap values are shown on nodes; branch lengths reflect estimated number of changes per site



P. mucidus and *P. submucidus*. The two latter species have thin-walled basidia and no sterile hymenial elements while basidia are slightly thick-walled in a senescent hymenium of *P. abiens* and hyphidia are present. Rather tough basidiocarps consisting of predominantly thick-walled hyphae and short, tardily thick-walled basidia of *P. abiens* are reminiscent of hydroid-semiporoid *Sistotremastrum* species, i.e., *S. aculeatum*, *S. aculeocrepitans*, and *S. denticulatum*. The only reliable anatomical traits differentiating them from *P. abiens* are their suburniform basidia and somewhat larger basidiospores. Moreover, no truly poroid species are so far known in *Sistotremastrum* s. str.

***Porpomyces abiens* Vlasák & Spirin, sp. nov.**—Figs. 5 and 6

MB 833939

Holotype. French Guiana. Remire-Montjoly: Lac du Rorota, rotten angiosperm wood, 23.VIII.2018 *Vlasák 1808/16** (H 7009714).

Etymology. *Abiens* (Lat., part. from *abeo*)—departing, deviating.

Basidiocarps effused, first soft-floccose, then rather tough, covering several cm, 0.2–1 mm thick. Hymenial surface pale cream-colored to yellowish or pale ochraceous, poroid, pores angular to sinuous, partly fusing together, strongly elongated on sloping substrate, 6–9 per mm, with thin, entire dissepiments. Margin white to pale cream-colored, fibrillose or compact, in some portions with thin, white, pronounced hyphal strands.

Hyphal structure monomitic; hyphae clamped. Subicular hyphae slightly to distinctly thick-walled, (2.2–) 2.7–7.3 (–7.8) μm in diam. ($n = 20/1$), interwoven or in subparallel bundles, often distinctly inflated (up to 10 μm); some hyphae encrusted by densely distributed thorn-like crystals or subglobose resinous globules. Tramal hyphae distinctly thick-walled, rather tightly arranged, interwoven to subparallel, occasionally anastomosing, partly glued together, (2.2–) 2.8–5.8 (–6.4) μm in diam. ($n = 40/2$), sometimes inflated near septa; some hyphae encrusted by densely distributed subglobose resinous globules. Subhymenial hyphae thin- to slightly thick-walled, mostly short-celled and distinctly inflated, (2.0–) 2.2–5.2 (–5.4) μm in diam. ($n = 40/2$). Rhomboid or prismatic crystals occasionally present among subicular or tramal hyphae, up to 15 μm in widest dimension, solitary or in large groups. Hyphidia rarely present, 10–12 \times 3.5–5 μm . Basidia subglobose to barrel-shaped or shortly clavate, 2–4-spored, (4.8–) 4.9–7.9 (–8.0) \times (3.4–) 3.7–4.8 (–5.0) μm ($n = 30/2$), senescent basidia slightly thick-walled. Basidioles subglobose to globose, 4–5 \times 4–4.5 μm . Basidiospores broadly cylindrical to ellipsoid, more rarely somewhat tapering to the apex (lacrymoid), (2.3–) 2.5–3.2 (–3.3) \times (1.6–) 1.7–2.2 (–2.3) μm ($n = 60/2$), $L =$

2.87–2.94, $W = 1.98$ –2.00, $Q = 1.46$ –1.47, contents homogeneous and CB (+).

Remarks. *Porpomyces abiens* has been detected in three localities in French Guiana. It seems to be a saprotrophic species occurring on wood remnants, various debris and soil.

Specimens examined. French Guiana. Remire-Montjoly: Lac du Rorota, hardwood, 23.VIII.2018 *Vlasák 1808/39** (H, JV). Roura: Natural Reserve of Kaw, Patawa Lodge, on soil between the roots of uprooted tree, 31.VIII.2018 *Vlasák 1808/146* (JV), Amazon Lodge, rotten hardwood log, 4.IX.2019 *Vlasák 1909/29** (JV).

***Sertulicium* Spirin, Volobuev & K.H. Larss., gen. nov.**

MB 833941

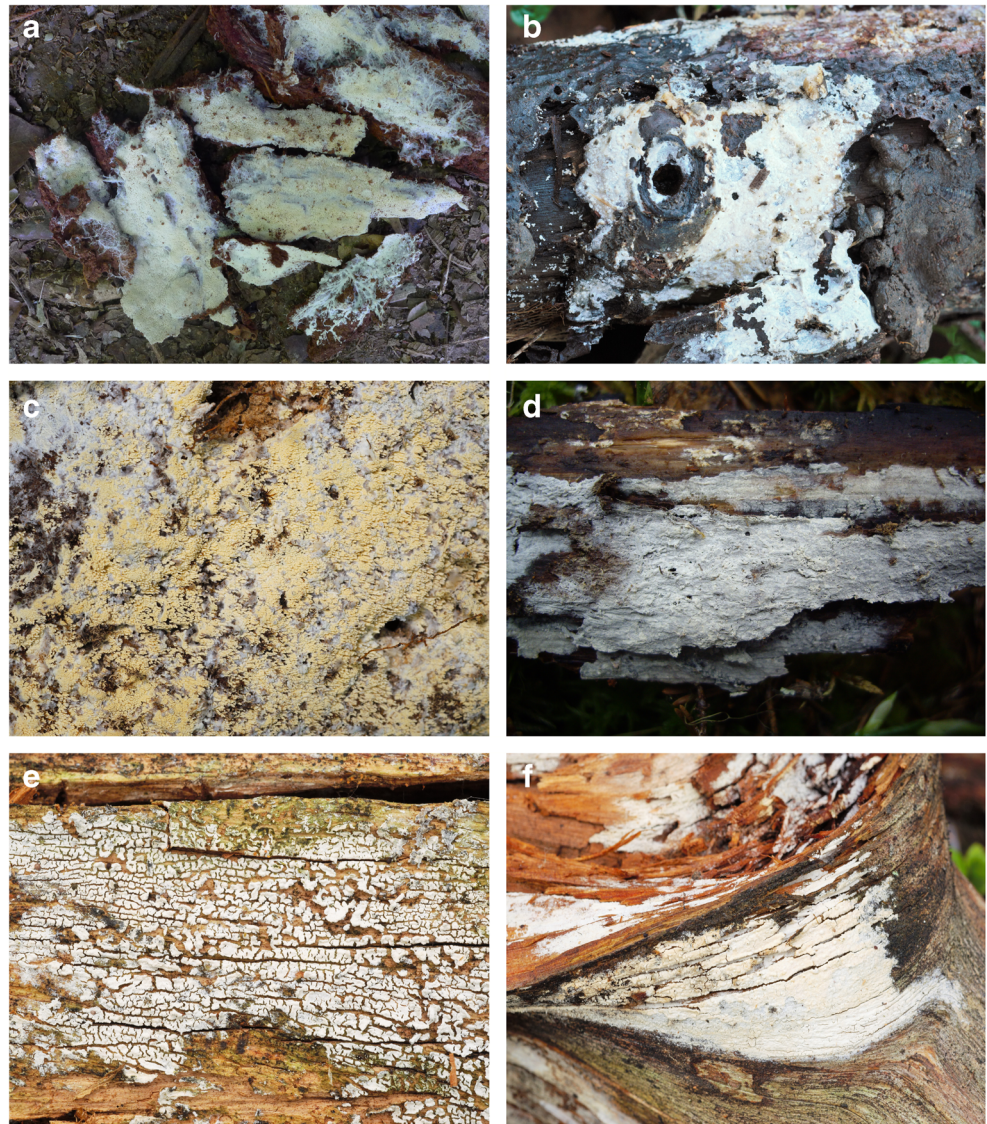
Etymology. *Sertulum* (Lat., noun)—a small chaplet.

Basidiocarps effused, very thin to rather thin (usually up to 0.1 mm thick), pruinose or waxy. Hymenophore smooth. Hyphal structure monomitic; all hyphae clamped, CB (+). Cystidia mostly absent, hyphidia rarely present, simple. Basidia clavate, with 4–6 sterigmata, normally not collapsing at the apex. Basidiospores thin-walled (but the wall distinct), narrowly ellipsoid to cylindrical, inamyloid, acyanophilous, contents homogeneous and CB (+). On rotten wood of deciduous trees and conifers.

Generic type. *Corticium niveocreum* Höhn. & Litsch.

Morphological differences between *Sertulicium* and *Sistotremastrum* s. str. cannot be easily grasped due to the extreme anatomical simplicity of their representatives. Nevertheless, some distinguishing characters are indicated here awaiting more in-depth analysis in the future. First, all but one *Sertulicium* spp. are extremely thin fungi consisting of a few subicular hyphae and the overlying subhymenium. The subicular hyphae are scattered; i.e., they do not produce hyphal strands so characteristic for most *Sistotremastrum* s. str. species studied by us. The only exception is *Sertulicium granuliferum*, having the most elaborate fructifications in the genus. Its basidiocarps start their development as radially arranged hyphal bundles becoming quickly covered by randomly arranged hyphae and finally indiscernible. In contrast, in *Sistotremastrum* s. str. subicular hyphal strands can be detected even in senescent basidiocarps if hyphae are not totally collapsed. Second, basidia in all *Sertulicium* spp. bear up to six sterigmata while at least some *Sistotremastrum* s. str. species are strictly four-sterigmatic. Moreover, apical parts of basidia in the latter genus often collapse, and hymenial cells of this kind dominate in well-developed and in senescent basidiocarps. Such apically collapsing basidia may occur in *Sertulicium* spp., too, but they are as a rule rare. Finally, all *Sertulicium* spp. have a smooth hymenial surface while about a half of the currently

Fig. 5 Basidiocarps of **a** *Porpomyces abiens* (holotype), **b** *Sertulicium granuliferum* (Miettinen 23459), **c** *Sistotremastrum aculeatum* (holotype), **d** *S. mendax* (Spirin 11208), **e** *S. suecicum* (Miettinen 13310), and **f** *S. suecicum* (senescent basidiocarps) (Miettinen 14548)



known *Sistotremastrum* spp. have a hydroid hymenophore.

Sertulicium chilense (Telleria, M. Dueñas & M.P. Martín) Spirin & Volobuev, **comb. nov.**

≡ *Sistotremastrum chilense* Telleria, M. Dueñas & M.P. Martín, Phytotaxa 158: 94, 2014 (as '*chilensis*').

MB 833942

Sistotremastrum chilense was described from Patagonia (Telleria et al. 2014) as a relative of *S. guttuliferum* (see under *Sertulicium granuliferum* below). From the latter species, it differs primarily in having long-celled subhymenial hyphae (vs. short-celled and somewhat inflated in *S. granuliferum*). DNA data confirm its placement in *Sertulicium*.

Sertulicium granuliferum (Hallenb.) Spirin & Volobuev, **comb. nov.**—Figs. 5 and 10

MB 833943

≡ *Trechispora granulifera* Hallenb., Iranian J. Plant Path. 14: 77, 1978. Holotype. Iran. Gilan: Asalem, *Parrotia persica*, 16.VII.1976 Hallenberg 1885 (GB 23442, studied).

= *Sistotremastrum guttuliferum* Melo, M. Dueñas, Telleria & M.P. Martín, Mycological Progress 12: 688, 2013. Holotype. Portugal. Madeira: Santana, decayed wood, 19.X.2006 Telleria 16566* (MA F82105, studied).

Basidiocarps effused, initially reticulate, soft-pellicular, then continuous, waxy, covering several cm, 0.05–0.1 mm thick. Hymenial surface first cream-colored, later yellowish to pale ochraceous, smooth or indistinctly tuberculate,

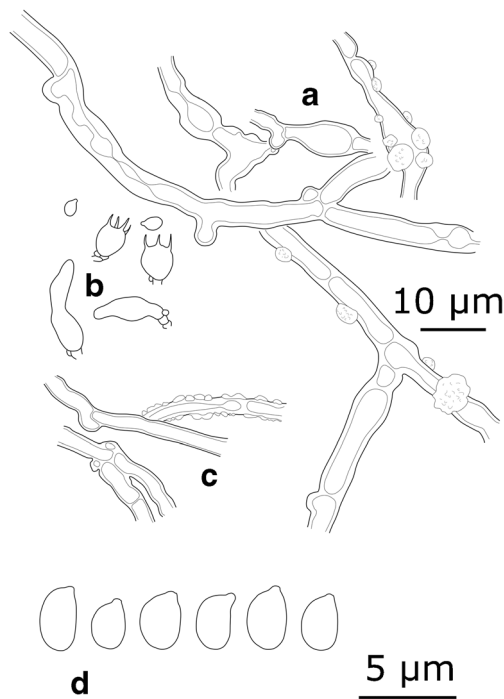


Fig. 6 Microscopic structures of *P. abiens* (holotype). **a** Subicular hyphae. **b** Hymenial cells. **c** Tramal hyphae. **d** Basidiospores

occasionally cracking in senescent basidiocarps. Margin first pruinose, later compact and rather sharply delimited. Small white patches spread at the marginal areas of young specimens, consisting of crystal agglomerations.

Hyphal structure monomitic; hyphae clamped, quickly collapsing. Subicular hyphae subparallel (juvenile specimens) to interwoven, thin- or slightly thick-walled, branched mostly at right angles, (3.0–) 3.1–6.5 (–6.9) µm in diam. ($n = 100/5$). Subhymenial hyphae thin-walled, densely arranged, some short-celled and slightly inflated, (2.2–) 2.3–4.2 (–4.6) µm in diam. ($n = 100/5$). Hyphidia occasionally present, embedded, simple, 3–4 µm in diam. Basidia clavate, 4–6-spored, gradually tapering to the basal part, (11.3–) 13.6–30.2 (–32.7) × (4.3–) 4.4–6.6 (–6.7) µm ($n = 100/5$). Basidioles clavate to bladder-shaped, (7.3–) 9.1–13.8 (–14.6) × (4.7–) 4.8–7.2 (–7.3) µm ($n = 30/2$). Basidiospores cylindrical, straight, rarely slightly curved, (4.8–) 4.9–7.3 (–8.0) × (2.3–) 2.5–3.3 (–3.5) µm ($n = 270/9$), $L = 5.40$ –6.76, $W = 2.78$ –3.07, $Q = 1.81$ –2.41.

Remarks. This species was introduced as a member of *Trechispora* due to some morphological similarity to the smooth-spored members of that genus (i.e., *Trechispora cohaerens* group) (Hallenberg 1978). Larsson (1992) found clavate basidia with 4–5 sterigmata in the type collection and thus pointed towards possible affinities with *Sistotremastrum*. After rechecking the type, as well as morphological and DNA studies of newly collected material, we conclude that *T. granulifera* is an older name for the species recently introduced as *Sistotremastrum guttuliferum* (Telleria et al. 2013a). Consequently, a new combination is proposed.

Sertulicium granuliferum differs from the similar-looking *S. niveocreameum* in having thicker and softer basidiocarps with a well-developed subicular layer and smaller basidiospores (Table 2).

Sertulicium granuliferum is widely distributed in temperate Eurasia. It seems to be not rare along the eastern coast of Canada and the USA, and its distribution in North America stretches to the central part of Mexico. All but one specimen came from wood remnants of deciduous trees, mostly from decayed wood but a few from still corticated, rather tough branches or logs. One specimen from Ethiopia is morphologically indistinguishable from other collections studied by us and therefore accepted here as *S. granuliferum*. Three other collections from Africa (Kenya and Tanzania) are microscopically identical to *S. granuliferum* but strikingly different macroscopically due to a well-developed, distinctly fibrillose margin. They may represent yet another species in this genus and thus treated as *Sertulicium* aff. *granuliferum* under Specimens examined. ITS regions of two published *Sistotremastrum* genomes (Nagy et al. 2016; *Sistotremastrum niveocreameum* LOAV000000000.1 and *Sistotremastrum suecicum* SRA:SRR1800527.68548550.2) appear to be conspecific and belong to *Sertulicium granuliferum*.

Specimens examined. Belgium. Oost-Vlaanderen: Kluisbergen, Kluisbos, deciduous wood, 20.II.2017 *Schouttetten* 17-021, 17-022 (GENT), 7.III.2017 *Schouttetten* 17-108 (GENT). Canada. Québec: Outaouais, Gatineau Nat. Park, angiosperm, 17.X.1967 *Eriksson* 9155 (GB), *Acer* sp., 17.X.1967 *Eriksson* 9177 (GB). Ethiopia. Shoa Prov.: Shashemene, dead hardwood twig, 18.VII.1990 *Ryvarden* 28557 (O). Germany. Nordrhein-Westfalen: Essen, Werden, deciduous wood, 6.I.2020 *Miettinen* 23459 (H). Italy. Lombardy: Varese, Valganna, San Gemolo, *Fraxinus excelsior*, 14.X.2019 *Spirin* 13657 (H). Mexico. Vera Cruz: Barranca de Pescado, angiosperm, 26.IX.1985 *Ryvarden* 23449 (O). Russia. Kemerovo Reg.: Novokuznetsk Dist., Kuzedeevo, *Betula* sp., 3.IX.2011 *Kotiranta* 28540 (H), *Salix caprea*, 4.IX.2011 *Kotiranta* 28597 (H). Leningrad Reg.: Boksitogorsk Dist., Chagoda, *Alnus incana*, 9.V.2018 *Spirin* 11877 (H). Nizhny Novgorod Reg.: Bogorodsk Dist., Krastelikh, *Tilia cordata*, 14.IX.2010 *Spirin* 3503 (H), *Q. robur*, 2.X.2015 *Spirin* 9898, 9899, 9910 (H), 17.VII.2016 *Spirin* 10310, 10313, 10315 (H), 5.V.2018 *Spirin* 11862 (H); Lukoyanov Dist., Panzelka, *Padus avium*, 17.VIII.2015 *Spirin* 9479 (H), 9.VIII.2016 *Spirin* 10577 (H), Razino, *Populus tremula*, 15.VIII.2015 *Spirin* 9296* (H), *Q. robur*, 10.VIII.2016 *Spirin* 10605 (H), *T. cordata*, 23.VII.2018 *Spirin* 12006 (H), Sanki, *Acer platanoides*, 20.VIII.2015 *Spirin* 9658 (H); Pavlovo Dist., Chudinovo, *P. tremula*, 3.X.2015 *Spirin* 9977 (H), 15.VII.2016 *Spirin* 10281, 10284 (H), *T. cordata*, 15.VII.2016 *Spirin* 10263 (H). Oryol Reg.: Krasnozorensky Dist., Malinovo, *Q. robur*, 7.X.2012 *Volobuev* (LE 314056*); Novoderevenkovsky

Table 2 Basidiospore dimensions of *Porpomyces* and *Sistotremastrum* s. lato species. Species name and basidiospore dimensions for a given taxon are presented in bold

Species/specimen	L'	L	W'	W	Q'	Q	n
<i>Porpomyces abiens</i>	(2.3) 2.5–3.2 (3.3)	2.91	(1.6) 1.7–2.2 (2.3)	1.99	(1.2) 1.3–1.7 (1.8)	1.47	60
holotype	(2.4) 2.5–3.2 (3.3)	2.94	(1.6) 1.7–2.2	2.00	(1.2) 1.3–1.7 (1.8)	1.47	30
Vlasák 1808/39	(2.3) 2.6–3.2 (3.3)	2.87	(1.7) 1.8–2.2 (2.3)	1.98	(1.3) 1.4–1.6 (1.7)	1.46	30
<i>Sertulicium granuliferum</i>	(4.8) 4.9–7.3 (8.0)	5.75	(2.3) 2.5–3.3 (3.5)	2.90	(1.5) 1.6–2.4 (2.5)	2.02	270
Kotiranta 26776	5.1–6.7 (6.8)	5.70	(2.6) 2.7–3.2 (3.3)	2.88	(1.6) 1.8–2.2 (2.3)	1.98	30
Larsson 12282	(5.0) 5.1–6.0 (6.1)	5.46	(2.7) 2.8–3.2 (3.3)	3.02	(1.6) 1.7–2.0 (2.1)	1.81	30
LE 299057	(4.8) 4.9–6.0 (6.1)	5.40	(2.5) 2.6–3.2	2.83	(1.6) 1.7–2.1 (2.2)	1.91	30
Miettinen 16083	(6.0) 6.1–7.3 (8.0)	6.76	(2.7) 2.8–3.3 (3.5)	3.07	(1.8) 2.0–2.4	2.21	30
Spirin 3503	(5.1) 5.2–6.2 (6.3)	5.82	(2.6) 2.7–3.1 (3.2)	2.85	(1.8) 1.9–2.3 (2.4)	2.04	30
Spirin 9296	5.2–6.5 (7.0)	5.95	(2.6) 2.7–3.2	2.87	(1.6) 1.8–2.4 (2.5)	2.41	30
Spirin 9898	(4.9) 5.0–6.1 (6.2)	5.46	(2.6) 2.7–3.2 (3.3)	2.86	(1.6) 1.7–2.1 (2.2)	1.92	30
Spirin 9977	(4.8) 5.1–6.6 (7.2)	5.63	(2.6) 2.7–3.3 (3.5)	2.97	(1.5) 1.6–2.2 (2.3)	1.91	30
Telleria 16566	(4.9) 5.0–6.1 (6.2)	5.56	(2.3) 2.5–3.1 (3.3)	2.78	(1.7) 1.8–2.2 (2.3)	2.00	30
<i>S. jacksonii</i>	(5.1) 5.2–8.2 (8.6)	6.60	(2.7) 2.8–4.1 (4.2)	3.29	(1.5) 1.6–2.6 (2.7)	2.02	170
holotype	(6.1) 6.2–8.0 (8.2)	7.13	(2.7) 2.8–3.2 (3.3)	2.99	2.2–2.6 (2.7)	2.39	20
Bernicchia 5464	(5.3) 5.7–7.3 (7.8)	6.52	(3.2) 3.3–4.1 (4.2)	3.66	(1.5) 1.6–2.1 (2.2)	1.79	30
Laurila 3256b	(5.2) 5.3–7.2	6.43	(2.7) 2.8–3.8 (4.0)	3.23	(1.8) 1.9–2.5 (2.6)	2.00	30
Miettinen 17141.2	(6.0) 6.1–8.2 (8.6)	6.96	3.0–3.4 (3.5)	3.21	(1.9) 2.0–2.5	2.17	30
Spirin 10425	(5.2) 5.6–7.4 (8.0)	6.49	(2.8) 2.9–3.8 (3.9)	3.30	(1.6) 1.7–2.3 (2.6)	1.97	30
Svantesson 699	(5.1) 5.2–6.8 (7.1)	6.08	(3.0) 3.1–3.9 (4.0)	3.36	(1.5) 1.6–1.9 (2.3)	1.81	30
<i>S. lateclavigerum</i>	(4.3) 4.6–7.2 (7.3)	5.57	(2.1) 2.2–3.2	2.71	(1.7) 1.8–2.4 (2.5)	2.07	60
holotype	(4.9) 5.0–7.2 (7.3)	5.82	(2.5) 2.6–3.2	2.84	(1.7) 1.8–2.3 (2.4)	2.05	30
Spirin 13457	(4.3) 4.6–6.2 (7.2)	5.32	(2.1) 2.2–3.0 (3.1)	2.57	(1.7) 1.8–2.4 (2.5)	2.09	30
<i>S. niveocreum</i>	(5.2) 5.5–8.6 (9.8)	6.86	(2.6) 2.7–4.2 (4.3)	3.25	(1.6) 1.7–2.4 (2.6)	2.12	120
holotype	(5.2) 5.5–7.8 (9.8)	6.62	(2.6) 2.7–3.6 (4.3)	3.12	(1.7) 1.9–2.4 (2.5)	2.12	30
Kotiranta 26267	(5.7) 6.0–7.4 (7.6)	6.55	(2.7) 2.8–3.4 (3.5)	3.08	(1.9) 2.0–2.3 (2.5)	2.13	30
Miettinen 14925.3	(6.0) 6.1–8.6 (9.2)	7.46	(3.0) 3.1–3.6 (3.8)	3.33	(1.7) 1.8–2.4 (2.6)	2.24	30
Söderholm 4050	(6.0) 6.1–7.7 (7.8)	6.79	(3.0) 3.1–4.2 (4.3)	3.46	(1.6) 1.7–2.2 (2.3)	1.97	30
<i>S. vernale</i>	(5.7) 5.8–8.2 (8.3)	6.96	(2.8) 2.9–3.8 (3.9)	3.30	(1.6) 1.8–2.5 (2.9)	2.12	60
holotype	(5.7) 6.1–8.2 (8.3)	7.02	(2.8) 2.9–3.8	3.28	(1.6) 1.8–2.5 (2.9)	2.15	30
Spirin 14244	(5.7) 5.8–8.0 (8.1)	6.90	(2.9) 3.0–3.8 (3.9)	3.31	(1.6) 1.8–2.3 (2.4)	2.09	30
<i>Sistotremastrum aculeatum</i>	(4.1) 4.2–6.2 (6.8)	5.03	(2.1) 2.2–3.1 (3.3)	2.58	(1.5) 1.6–2.4 (2.7)	1.97	120
holotype	(4.1) 4.2–6.2 (6.8)	5.07	(2.2) 2.3–3.1 (3.3)	2.65	(1.5–)1.6–2.4 (2.7)	1.94	60
Dunaev KUN1105d	4.2–5.3 (5.4)	4.98	(2.1) 2.2–3.0 (3.1)	2.60	(1.6–)1.7–2.4 (2.5)	1.94	30
Miettinen 10380.1	(4.2) 4.3–6.1 (6.2)	5.05	(2.2) 2.3–2.8 (3.0)	2.50	1.8–2.2 (2.3)	2.03	30
<i>S. aculeocrepitans</i>	(4.0) 4.1–5.6 (5.8)	4.77	(2.1) 2.2–2.8 (2.9)	2.33	1.8–2.5 (2.6)	2.06	60
Larsson 16097	(4.4) 4.5–5.6 (5.8)	5.10	(2.1) 2.2–2.8 (2.9)	2.35	(1.8) 1.9–2.5 (2.6)	2.19	30
Larsson 16478	(4.0) 4.1–5.0 (5.1)	4.44	(2.1) 2.2–2.6 (2.7)	2.30	1.8–2.1 (2.2)	1.93	30
<i>S. confusum</i>	(5.3) 5.7–7.6 (7.9)	6.56	2.9–3.9 (4.0)	3.24	(1.7) 1.8–2.3 (2.6)	2.03	60
holotype	(5.3) 5.7–7.3 (7.8)	6.54	2.9–3.4 (3.8)	3.16	(1.7) 1.8–2.3 (2.6)	2.07	30
Larsson 16004	(5.3) 5.7–7.6 (7.9)	6.57	3.0–3.9 (4.0)	3.32	(1.7) 1.8–2.3 (2.4)	1.99	30
<i>S. denticulatum</i>	(3.9) 4.0–5.3 (5.4)	4.51	(2.0) 2.1–3.0 (3.1)	2.48	(1.5) 1.6–2.3 (2.4)	1.83	30
holotype	(3.9) 4.0–5.3 (5.4)	4.51	(2.0) 2.1–3.0 (3.1)	2.48	(1.5) 1.6–2.3 (2.4)	1.83	30
<i>S. fibrillosum</i>	(4.0) 4.2–5.2 (5.6)	4.72	(2.1) 2.2–2.9 (3.0)	2.53	(1.6–)1.7–2.1 (2.2)	1.88	20
Larsson 16988	(4.0) 4.2–5.2 (5.6)	4.72	(2.1) 2.2–2.9 (3.0)	2.53	(1.6–)1.7–2.1 (2.2)	1.88	20
<i>S. geminum</i>	(3.8) 3.9–5.2 (5.5)	4.28	(1.9) 2.0–2.6 (2.9)	2.24	(1.7) 1.8–2.2 (2.3)	1.92	30
holotype	(3.8) 3.9–5.2 (5.5)	4.28	(1.9) 2.0–2.6 (2.9)	2.24	(1.7) 1.8–2.2 (2.3)	1.92	30
<i>S. induratum</i>	(4.0) 4.1–5.3 (5.8)	4.55	(2.5) 2.6–3.1 (3.2)	2.83	1.4–1.8 (1.9)	1.62	60

Table 2 (continued)

Species/specimen	L'	L	W'	W	Q'	Q	n
holotype	(4.1) 4.2–5.3 (5.8)	4.73	(2.5) 2.6–3.0	2.79	(1.5) 1.6–1.8 (1.9)	1.70	30
Spirin 8804	(4.0) 4.1–5.0 (5.1)	4.37	(2.5) 2.6–3.1 (3.2)	2.86	1.4–1.8 (1.9)	1.53	30
<i>S. mendax</i>	(4.0) 4.2–7.0 (7.2)	5.27	(2.4) 2.5–3.5 (3.6)	2.93	(1.3) 1.4–2.1 (2.3)	1.80	180
holotype	(4.7) 4.8–6.2	5.60	(2.7) 2.9–3.5 (3.6)	3.09	(1.6) 1.7–2.1 (2.3)	1.82	30
Larsson 12022	(4.6) 4.8–5.6	5.12	(2.6) 2.7–3.0 (3.1)	2.85	(1.6) 1.7–1.9	1.80	30
Nordén 9579	(4.7) 4.8–5.8 (6.0)	5.33	(2.6) 2.7–3.1 (3.2)	2.93	(1.6) 1.7–2.0 (2.1)	1.82	30
Spirin 10060	(5.0) 5.1–7.0 (7.2)	5.71	(2.6) 2.7–3.2 (3.3)	2.93	1.7–2.3 (2.4)	1.95	30
Spirin 10107	(4.2) 4.3–5.9 (6.1)	5.11	(2.7) 2.8–3.2 (3.5)	3.00	(1.3) 1.4–1.9 (2.0)	1.71	30
Vlasák 1110/14.1	(4.0) 4.2–5.4 (5.5)	4.75	(2.4) 2.5–3.0	2.76	(1.4) 1.5–1.9	1.72	30
<i>S. rigidum</i>	(3.8) 4.0–5.1 (5.2)	4.44	2.0–2.5 (2.7)	2.22	(1.6) 1.7–2.2 (2.3)	2.01	30
holotype	(3.8) 4.0–5.1 (5.2)	4.44	2.0–2.5 (2.7)	2.22	(1.6) 1.7–2.2 (2.3)	2.01	30
<i>S. suecicum</i>	(4.1) 4.2–6.2 (6.3)	4.90	(2.1) 2.2–3.1 (3.2)	2.53	(1.5) 1.6–2.4 (2.5)	1.93	180
holotype	(4.8) 5.1–6.2 (6.3)	5.55	(2.4) 2.5–3.1 (3.2)	2.80	(1.7) 1.8–2.2 (2.3)	1.99	30
Kunttu 5959	(4.1) 4.2–5.2	4.70	2.2–2.8 (2.9)	2.52	(1.6) 1.7–2.1 (2.2)	1.88	30
Larsson 11849	(4.1) 4.2–5.7 (5.8)	4.91	(2.1) 2.2–2.9	2.42	(1.7) 1.8–2.4 (2.5)	2.05	30
Miettinen 11044	(4.5) 4.6–5.9 (6.1)	5.12	2.2–2.9 (3.0)	2.50	(1.7) 1.8–2.3 (2.5)	2.06	30
Miettinen 14550.1	(4.1) 4.2–5.1 (5.3)	4.60	2.2–2.9 (3.0)	2.52	(1.5) 1.6–2.0 (2.1)	1.83	30
Spirin 8932	(4.1) 4.2–5.3 (5.8)	4.56	(2.1) 2.2–2.9 (3.0)	2.42	(1.7) 1.8–2.1 (2.2)	1.89	30
<i>S. vigilans</i>	(6.1) 6.3–11.2 (11.4)	8.68	(3.0) 3.1–5.4 (6.2)	4.35	(1.6) 1.7–2.5 (2.6)	2.01	150
holotype	(7.0) 7.2–9.1 (9.4)	8.09	(3.9) 4.0–5.2 (5.8)	4.49	(1.6) 1.7–2.0 (2.1)	1.81	30
Eriksson 9106	(8.0) 8.3–10.5 (10.6)	9.34	(3.8) 4.0–5.0 (5.1)	4.44	(1.8) 1.9–2.2 (2.3)	2.11	30
Fonneland 2011-77	(8.1) 8.8–11.2 (11.4)	9.95	(4.0) 4.1–5.1 (5.2)	4.50	(1.8) 1.9–2.5 (2.6)	2.22	30
Häyrén 1934	(7.7) 7.8–10.3 (10.9)	8.61	(4.1) 4.2–5.4 (6.2)	4.69	(1.7) 1.8–2.0 (2.1)	1.84	30
Spirin 8778	(6.1) 6.3–8.8 (8.9)	7.43	(3.0) 3.1–4.3 (4.5)	3.64	(1.7) 1.8–2.3 (2.4)	2.05	30

Dist., Mokhovoe, *Q. robur*, 24.VII.2011 Volobuev (LE 314057*); Sverdlovsky Dist., Maryevka, *Betula pendula*, 1.IX.2012 Volobuev (LE 314055*); Uritsky Dist., Naryshkino, *B. pendula*, 6.X.2012 Volobuev (LE 292194*), *Q. robur*, 25.VI.2011 Volobuev (LE 314054*), 6.X.2012 Volobuev (LE 314053*). Primorie Reg.: Ternei Dist., Maisa, hardwood stump, 9.IX.1990 Larsson 7182 (GB), *Alnus* sp. (?), 12.IX.1990 Larsson 7447 (GB), Blagodatnoe, *Alnus* sp., 23.IX.1990 Larsson 8140 (GB). Tuva: Turgen, *Larix sibirica* (charred corticated log), 23.VIII.2014 Kotiranta 26776* (H). Spain. Burgos: Ordun, *Fagus sylvatica*, 12.XI.1977 Ryvardeen 15329 (O, H). Santander: Cavadonga, *Sambucus*, 14.IX.1977 Ryvardeen 15365 (O, H). Ukraine. Donetsk Reg.: Slovyansk Dist., Pyskunkivka, *Salix alba*, 20.XI.2010 Akulov (CWU 4704*). USA. Massachusetts: Hampshire Co., South Hadley, decayed wood, 4.V.2013 Miettinen 16083* (H); Worcester Co., Holden, cut bole, 26.IX.2011 Miettinen 14813.2* (H), Worcester, ?*Acer saccharum*, 8.IX.2013 Miettinen 16834 (H), *Populus tremuloides*, 5.X.2013 Miettinen 17267.4 (H), *Quercus* sp., 6.X.2013 Miettinen 17272* (H), *Acer* sp., 26.X.2014 Miettinen 19035 (H). Michigan: Barry Co., Yankee Springs, *Quercus* sp., 3.IX.1955 Cain (H ex TRTC 31857). Tennessee: Cocke Co., Cosby, decayed wood, 13.VII.2004 K.H. Larsson

12198 (GB), 14.VII.2004 K.H. Larsson 12216 (GB), 17.VII.2004 K.H. Larsson 12282*, 12297* (GB).

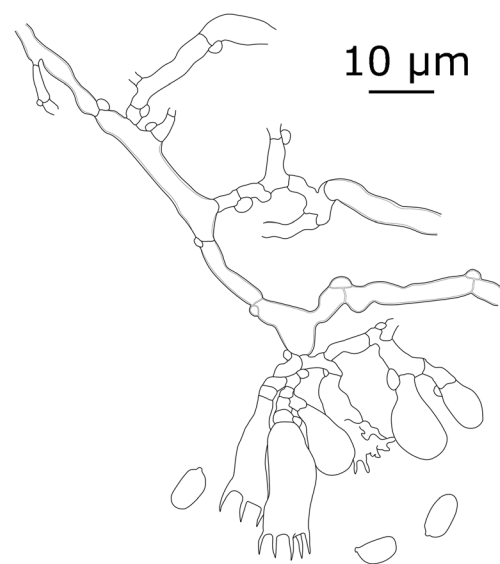


Fig. 7 Microscopic structures of *S. jacksonii* (Svantesson 699): hyphae, hymenial cells, and basidiospores

Sertulicium jacksonii (Liberta) Spirin & K.H. Larss., **comb. nov.**—Figs. 7 and 10

MB 833944

≡ *Paullicorticium jacksonii* Liberta, Brittonia 14: 223, 1962. Holotype. Canada. Ontario: York, Maple, decayed wood, 11.IX.1943 Jackson (TRTC 18722, studied).

Basidiocarps effused, pruinose, hardly visible by the naked eye, 0.02–0.03 mm thick. Hymenial surface whitish, smooth or porulose. Margin indistinct.

Hyphal structure monomitic; hyphae clamped. Subicular hyphae sparse, randomly arranged, slightly to moderately thick-walled, branched mostly at sharp angles, often anastomosing, (2.2–) 2.3–6.4 (–6.8) μm in diam. ($n = 68/5$). Subhymenial hyphae thin- or slightly thick-walled, densely and randomly arranged, (2.0–) 2.1–3.8 (–4.0) μm in diam. ($n = 100/5$). Hyphidia occasionally present, simple, slightly projecting, 3–4 μm in diam. Basidia clavate, 4–6-spored, gradually tapering to the basal part, (11.6–) 12.2–23.8 (–24.7) \times (5.8–) 5.9–9.3 (–9.4) μm ($n = 72/4$), a few pleural basidia seen. Basidioles broadly clavate to subglobose, (8.2–) 8.3–13.2 (–13.3) \times (4.7–) 5.7–9.2 (–9.3) μm ($n = 35/4$). Basidiospores cylindrical to narrowly ellipsoid, straight or slightly curved, longest spores slightly tapering to the distal end, (5.1–) 5.2–8.2 (–8.6) \times (2.7–) 2.8–4.1 (–4.2) μm ($n = 170/6$), $L = 6.08$ –7.13, $W = 2.99$ –3.66, $Q = 1.79$ –2.39.

Remarks. *Paullicorticium jacksonii* was described from Canada but later rarely treated as a species of its own. Eriksson et al. (1978) placed *P. jacksonii* among the synonyms of *Sistotremastrum niveocreum*, and the species has been interpreted that way up to the present moment. Our data confirm that *P. jacksonii* is a good species belonging to *Sertulicium*. In addition to the type collection, we studied specimens from the USA and Europe.

Sertulicium jacksonii belongs to a difficult complex of extremely thin, conifer-dwelling species hardly distinguishable via morphological traits. Among them, *S. jacksonii* can be recognized by its wide subicular hyphae with variably thickened walls. Two similar-looking species, *Sertulicium vernale* and *S. lateclavigerum*, have narrower subicular hyphae than those of *S. jacksonii*. Additionally, *S. lateclavigerum* possesses narrower basidiospores than in the other two species (Table 2). However, the differences listed here were detected from a very restricted set of specimens. Further studies with broader sampling may improve our observations considerably. *Paullicorticium ansatum* Liberta, yet another morphologically similar although phylogenetically unrelated species, may be easily separated from *Sertulicium* spp. due to ansiform (loop-like) clamps and basidia with up to eight sterigmata.

Specimens examined. Italy. Trentino—Alto Adige: Bolzano, *Picea abies*, 30.VIII.1990 Bernicchia 5464 (O ex HUBO). Norway. Oppland: Jevnaker, *P. abies*, 4.X.2011 Svantesson 699* (O). Russia. Karelia:

Medvezhiegorsk Dist., Krivozero, *P. abies*, 19.VI.1942 Laurila 3256b (H). Leningrad Reg.: Boksitogorsk Dist., Kolp', *P. abies*, 28.VII.2016 Spirin 10425* (H). USA. New York: Essex Co., Arbutus Lake, *Abies* sp., 22.IX.2013 Miettinen 17141* (H).

Sertulicium lateclavigerum (Boidin & Gilles) Spirin & Viner, **comb. nov.**—Fig. 8

MB 833945

≡ *Sistotremastrum lateclavigerum* Boidin & Gilles, Bull. Soc. Mycol. France 110: 217, 1994. Holotype. France. Landes: Tartas, *Pinus pinaster* (fallen branch), 17.VI.1988 Gilles 765* (LY-JB 13467, studied).

Basidiocarps effused, pruinose, rather loose or compact, covering a few cm, 0.02–0.05 mm thick. Hymenial surface cream-colored, smooth or porulose. Margin indistinct.

Hyphal structure monomitic; hyphae clamped. Subicular hyphae sparse, randomly arranged, thin- or only slightly thick-walled, branched mostly at sharp angles, often anastomosing, (2.2–) 2.3–4.3 (–4.9) μm in diam. ($n = 40/2$). Subhymenial hyphae thin-walled (wall distinct), rather loosely and randomly arranged, (2.0–) 2.1–3.7 (–2.8) μm in diam. ($n = 40/2$). Cystidia occasionally present, thin-walled, easily collapsing, broadly clavate or subglobose, 19–33 \times 9–16.5 μm . Hyphidia occasionally present, embedded, simple, 2–2.5 μm in diam. Basidia clavate, 4–6-spored, gradually tapering to the basal part, (9.3–) 9.9–20.7 (–21.2) \times (4.4–) 4.8–9.3 (–11.1) μm ($n = 40/2$), a few pleural basidia seen. Basidioles broadly clavate to bladder-shaped or subglobose, (7.3–) 7.9–10.2 (–10.3) \times (4.4–) 5.4–6.6 (–7.9) μm ($n = 20/1$). Basidiospores cylindrical to subfusiform, sometimes slightly curved, (4.3–)

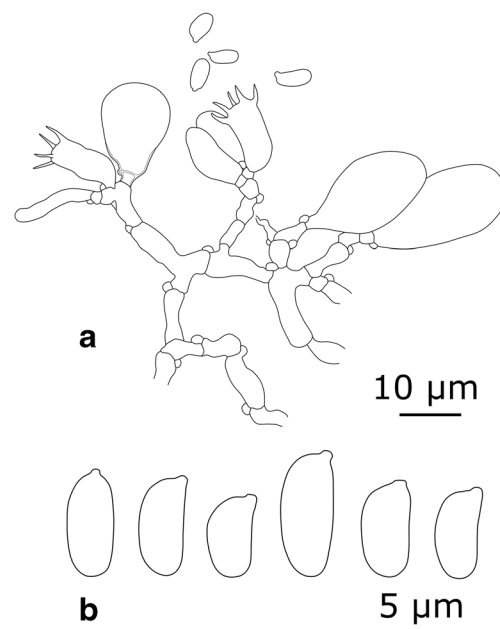


Fig. 8 Microscopic structures of *S. lateclavigerum* (holotype). **a** Hyphae and hymenial cells. **b** Basidiospores

4.6–7.2 (–7.3) \times (2.1–) 2.2–3.2 μm ($n = 60/2$), $L = 5.32$ –5.82, $W = 2.57$ –2.84, $Q = 2.05$ –2.09.

Remarks. *Sistotremastrum lateclavigerum* was described based on a single collection from France. The diagnostic features treated in the protologue were the bubble-like cystidia, basidia with up to 8 sterigmata and rather narrow and slightly curved basidiospores (Boidin and Gilles 1994). Sequences obtained from the holotype were published by Gruhn et al. (2018). They indicate that the species belongs to *Sertulicium* and it is not closely related to the similarly looking *S. jacksonii* and *S. vernale*. Morphological differences of *S. lateclavigerum* versus the two latter species are presented under *S. jacksonii*. *Paullicorticium allantosporum* J. Erikss. has basidiospores similar to *S. lateclavigerum*, (5.2–) 5.3–7.2 \times (2.3–) 2.4–3.2 (–3.3) μm ($n = 30/1$), $L = 6.10$, $W = 2.79$, $Q = 2.20$, but they are more clearly curved and have large oil droplets in the cytoplasm. Moreover, basidia of *P. allantosporum* are 6–8-sterigmatic, with a clearly contracted and more or less sinuous basal part.

We studied the holotype of *S. lateclavigerum* and observed basidia with 4–6 sterigmata only. In our opinion, the presence of globose or broadly clavate cystidia-like cells is insufficient as a single diagnostic character in *Sistotremastrum* s. l. We observed hymenial cells of the same shape in one collection of *S. jacksonii* (Laurila 3256b), in *S. niveocreum* (Van Autgaerden S-20), as well as in three representatives of *Sistotremastrum* s. str. (holotype of *S. geminum* and two senescent collections of *S. mendax* and *S. vigilans*, respectively). On the other hand, another collection of *S. lateclavigerum* studied by us (Spirin 13457) is totally devoid of cystidia-like cells.

So far, *S. lateclavigerum* has been collected in France and Slovenia. However, environmental sequences in GenBank and UNITE point to a broader distribution area of this species. One of them came from Vietnam (MF942562) while the second (UDB051695) is from Estonia.

Specimens examined. Slovenia. Gorenjska: Mojstrana, Triglavsko Bistrica, *P. abies*, 28.IX.2019 Spirin 13457* (H).

Sertulicium niveocreum (Höhn. & Litsch.) Spirin & K.H. Larss., **comb. nov.**—Fig. 10

MB 833946

≡ *Corticium niveocreum* Höhn. & Litsch., Sitzungsber. Kaiserl. Akad. Naturwiss. Mat.-Naturwiss. Klasse I 117:1117, 1908. Holotype. Austria. Niederösterreich: Wien-Umgebung, Unter-Tullnerbach, *Fagus sylvatica*, 11.III.1905 Höhnel (W 16251, studied).

= *Galzinia vesana* Boidin & Gilles, Bull. Soc. Mycol. France 106: 158, 1990. Holotype. France. Landes: St. Perdon, *Quercus pedunculata*, 2.III.1989 Gilles 828 (LY-JB 13675, studied).

Basidiocarps effused, initially pruinose, later compact, covering several cm, 0.03–0.1 mm thick. Hymenial surface

whitish to cream-colored or pale ochraceous, smooth or porulose. Margin first pruinose, later compact and sharply delimited.

Hyphal structure monomitic; hyphae clamped. Subicular hyphae interwoven to subparallel, slightly thick-walled, branched mostly at right angles, (3.6–) 3.7–5.3 (–5.6) μm in diam. ($n = 40/2$); some hyphae inflated at septa up to 6 μm in diam. Subhymenial hyphae thin- or slightly thick-walled, densely arranged, some short-celled and slightly inflated, (2.2–) 2.3–4.3 (–4.5) μm in diam. ($n = 80/4$). Hyphidia occasionally present, embedded, simple, 2.5–4.5 μm in diam. Basidia clavate, 4–6-spored, gradually tapering to the basal part, (10.7–) 10.8–24.3 (–26.7) \times (5.1–) 5.2–8.3 (–8.4) μm ($n = 60/3$), some basidia slightly thick-walled at the basal part. Basidioles broadly clavate to bladder-shaped or subglobose, (7.2–) 8.4–14.3 (–14.7) \times (4.1–) 4.2–8.3 (–8.4) μm ($n = 80/4$). Basidiospores cylindrical, straight or slightly curved, (5.2–) 5.5–8.6 (–9.8) \times (2.6–) 2.7–4.2 (–4.3) μm ($n = 120/4$), $L = 6.55$ –7.46, $W = 3.08$ –3.46, $Q = 1.97$ –2.24.

Remarks. *Sertulicium niveocreum* is distributed in temperate–boreal Eurasia. It inhabits angiosperm hosts, mainly still attached or recently fallen branches but it may occur also on rather rotten wood. For a long time, *S. niveocreum* has been mixed up with another angiosperm-dwelling species, *S. granuliferum*; their differences are discussed under the latter species. We studied the type material and a recent collection of *Galzinia vesana* from France; in our opinion, it is merely a monstrous form of *S. niveocreum* with unusually long basidia and basidiospores, as well as occasionally branched sterigmata. DNA data from recently collected specimens confirmed this synonymy. While introducing *G. vesana*, Boidin and Gilles (1990) referred to a description and illustration of *Galzinia* sp. from Sweden (Eriksson and Ryvarden 1975: 398) as another possible record of that species. We re-checked the specimen illustrated by Eriksson and concluded that it belongs to *Sertulicium*, being morphologically most similar to *S. niveocreum*. However, its exact identity is still uncertain, and it is therefore treated as *Sertulicium* sp. 2 under Specimens examined.

Some specimens of *S. niveocreum* collected in the winter or spring are dominated by four-sterigmatic basidia. Only a few 5–6-sterigmatic basidia were detected in such specimens after meticulous study. Moreover, these collections usually have longer basidiospores than in most other specimens of *S. niveocreum* (in particular, reaching 14.5 μm long in specimen Van Autgaerden S-20) and thus can be mixed up with *Sistotremastrum vigilans*. The latter species, introduced below, has constantly four-sterigmatic basidia and it occurs exclusively on coniferous hosts while *S. niveocreum* seems to be restricted to angiosperms.

Specimens examined. Belgium. Antwerpen: Zoersel, Zoerselbos, deciduous wood, 16.V.2019 Van Autgaerden

S-20* (GENT). Oost-Vlaanderen: Kluisbergen, Kluisbos, deciduous wood, 20.II.2017 *Schoutteten 17-001* (GENT), 13.III.2017 *Schoutteten 17-202* (GENT). Finland. Varsinais-Suomi: Lohja, Tamminiemi, *Corylus avellana*, 5.XI.2020 *Spirin 14241* (H). Uusimaa: Helsinki, Veräjämäki, *Sorbus aucuparia* (?), 19.X.2011 *Miettinen 14925.3** (H). Etelä-Häme: Kangasala, Lorunkorpi, *Salix* sp., 23.III.2009 *Söderholm 4050** (H). France. Haut-Rhin: Sainte-Marie-aux-Mines, Freland, Le Limbach, *F. excelsior*, 12.X.2019 *Spirin 13566* (H). Pyrénées-Orientales: Argelès-sur-Mer, angiosperm (twigs), 2.XI.2008 *K.H. Larsson 13727** (GB). Italy. Emilia-Romagna: Bologna, *Quercus* sp., 3.XI.1983 *Bernicchia 1849* (O ex HUBO), 16.X.1984 *Ryvarden 22225* (O). Sardinia: Cagliari, 17.III.2003 *Arras 897* (H ex HUBO); Nuoro, *Quercus* sp., 30.III.2010 *Bernicchia 8593* (O ex HUBO). Russia. Krasnoyarsk Reg.: Turukhansk Dist., Bor, *Alnus sibirica*, 16.VIII.2013 *Kotiranta 26267** (H). Sakha Rep.: Nizhnekolymsk Dist., Tsherski, *Alnus fruticosa*, 18.VIII.1972 *Parmasto* (H ex TAAM 56632). Spain. Canary Islands: Tenerife, Las Mercedes, decayed wood, 18.I.1974 *Ryvarden 12639* (O, H). United Kingdom. England: Hampshire, New Forest Nat. Park, *F. sylvatica*, 5.II.1994 *Legon* (GB).

***Sertulicium vernale* Spirin & Volobuev, sp. nov.**—Fig. 9 MB 833947

Holotype. Finland. Etelä-Häme: Kangasala, Pikku-Salmus, 61.4138° 23.9275° ±300 m, moist depression in rather old *Picea* forest, on a fallen, decorticated *Picea abies*, 10.V.2007 *Söderholm 3886** (H 6003442, isotype—LE).

Etymology. *Vernalis* (Lat., adj.)—vernal, occurring in the spring.

Basidiocarps effused, pruinose, rather loose, covering a few cm, 0.02–0.04 mm thick. Hymenial surface greyish, smooth or porulose. Margin indistinct.

Hyphal structure monomitic; hyphae clamped. Subicular hyphae sparse, randomly arranged, slightly to moderately thick-walled, branched mostly at sharp angles, often anastomosing, (2.2–) 2.3–4.8 (–5.2) µm in diam. (n = 40/2). Subhymenial hyphae thin-walled (wall usually distinct), rather tightly and randomly arranged, (2.0–) 2.1–4.1 (–4.2) µm in diam. (n = 40/2), sometimes bearing bubble-like cells, 10–12 × 8–11 µm. Hyphidia rare, embedded, simple, 2.5–3.5 µm in diam. Basidia clavate, 4–6-spored, gradually tapering to the basal part, (11.2–) 11.8–27.4 (–37.2) × (5.0–) 5.1–7.2 (–7.3) µm (n = 44/2). Basidioles broadly clavate to bladder-shaped or subglobose, (6.0–) 6.9–10.2 (–10.8) × (4.4–) 5.2–6.8 (–6.9) µm (n = 20/1). Basidiospores narrowly ellipsoid to broadly cylindrical, sometimes slightly curved, (5.7–) 5.8–8.2 (–8.3) × (2.8–) 2.9–3.8 (–3.9) µm (n = 60/2), L = 6.90–7.02, W = 3.28–3.31, Q = 2.09–2.15.

Remarks. *Sertulicium vernale* is morphologically very similar to *S. jacksonii*, and it can be differentiated from the latter

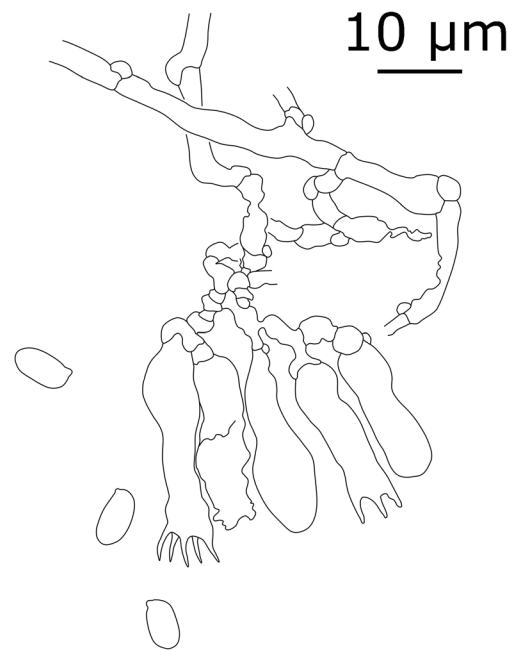


Fig. 9 Microscopic structures of *S. vernale* (holotype): hyphae, hymenial cells, and basidiospores

species by its narrower subicular hyphae and longer basidia. The species is so far known from two localities in Finland and two environmental sequence from Sweden (AY805625—Menkis et al. 2004) and CO, USA (JX136434—Huffman et al. 2013). The North American sequence is slightly different (2 bp) from the two European ones, and it may still belong to an undescribed sibling species.

Specimens examined. Finland. Varsinais-Suomi: Lohja, Tamminiemi, *P. abies*, 5.XI.2020 *Spirin 14244* (H).

***Sistotremastrum* J. Erikss., Symb. Bot. Upsalienses 16 (1): 62, 1958.**

Basidiocarps effused, thin to rather substantial (up to 0.2 mm thick), pruinose to waxy-compact. Hymenophore smooth, warted or odontoid-semiporoid. Hyphal structure monomitic; all hyphae clamped, CB (+), easily collapsing, in odontoid species with a more or less clear difference between trama and subhymenium. Cystidia absent in all but one species, hyphidia present, simple or accidentally branched. Basidia clavate or suburniform, with 2–4 or 4–6 sterigmata, usually collapsing at the apex, senescent basidia often slightly thick-walled at the base. Basidiospores thin-walled (but the wall distinct), narrowly ellipsoid or ovoid to cylindrical, inamyloid, acyanophilous, contents homogeneous and CB (+). On rotten wood of deciduous trees and conifers.

Generic type. *Sistotremastrum suecicum* Litsch. ex J. Erikss.

The genus is redefined here to exclude members of the *Sistotremastrum niveocremaeum* complex subsumed under *Sertulicium*. Differences between the two genera are discussed above. Extending *Sistotremastrum* s. str. with a number of four-

sterigmatic taxa makes it difficult to delimit the genus from *Brevicellicium* and from smooth-spored *Trechispora* species. *Trechispora* differs by generally soft basidiocarps, short, often almost isodiametric subhymenial cells, and presence of ampullate septa on subicular hyphae. *Brevicellicium* species have similar subhymenial cells as in *Trechispora* but lack ampullate septa in the subiculum, which is poorly developed. Moreover, basidia in the four-sterigmatic *Sistotremastrum* spp. are more or less clearly suburniform, i.e., slightly constricted at the middle and narrowed at the apical part (except *S. vigilans* with exceptionally long, clavate-utriform basidia) while they are barrel-shaped or short-clavate in *Brevicellicium* and *Trechispora*.

All but one four-sterigmatic representatives of the genus treated below are distributed in tropical areas. However, two unnamed sequences (CMH343, GenBank KF800434; 6S1.10.S04, GenBank EF619877) originated from the eastern part of the USA (Fig. 4). This could point towards more undescribed taxa in the four-sterigmatic species complex distributed in geographic regions with a cooler climate.

***Sistotremastrum aculeatum* Miettinen & Viner, sp. nov.**—Fig. 10

MB 833948

Holotype. Indonesia. Sumatera Barat: Kab. Lima Pulu Kota, Gunung Sago, -0.30585° 100.67762°, alt. 1400 m, lower montane rainforest, on a fallen log of dicot (decay stage 3/5), 25.VII.2009 Miettinen 13799.1* (ANDA—holotype, H 7009306—isotype).

Etymology. *Aculeatus* (Lat., adj.)—aculeate.

Basidiocarps effused, first soft-floccose, then rather tough, covering several to tens of cm, 0.1–0.2 mm thick. Hymenial

surface pale cream-colored to pale ochraceous, first warted, then odontoid, with irregularly arranged, sharp-pointed, partly fusing spines, 2–3 per mm, up to 2 mm long, or semiporoid, with shallow pores 0.5–1 per mm. Margin white to pale ochraceous, compact, in some portions with thin, white, pronounced hyphal strands. Smooth areas between spines fertile.

Hyphal structure monomitic; hyphae clamped. Subicular hyphae unevenly thick-walled, (2.8–) 2.9–5.1 (–5.2) μm in diam. ($n = 40/2$), often in subparallel bundles. Tramal hyphae distinctly thick-walled, rather tightly arranged, interwoven to subparallel, sometimes glued together, (2.7–) 3.0–5.6 (–6.2) μm in diam. ($n = 40/2$). Subhymenial hyphae slightly thick-walled, short-celled, (2.3–) 2.4–4.2 (–4.6) μm in diam. ($n = 40/2$), rarely inflated up to 6 μm . Rhomboid or prismatic crystals occasionally present among subicular or tramal hyphae, up to 9 μm in widest dimension, solitary or in large groups. Hyphidia occasionally present, embedded or slightly projecting, simple, 2–3 μm in diam. Basidia suburniform, 2–4-spored, (7.3–) 7.7–12.2 (–12.3) \times (3.3–) 3.4–4.5 (–5.2) μm ($n = 30/2$), senescent basidia slightly thick-walled at the basal part. Basidioles clavate to bladder-shaped, 5.4–8.2 \times 3.2–4.3 μm ($n = 10/1$). Basidiospores cylindrical to narrowly ovoid, straight, (4.1–) 4.2–6.2 (–6.8) \times (2.1–) 2.2–3.1 (–3.3) μm ($n = 120/3$), $L = 4.98$ –5.07, $W = 2.50$ –2.65, $Q = 1.94$ –2.03.

Remarks. *Sistotremastrum aculeatum* seems to be widely distributed in South-East Asia from China to Sumatra. Morphologically, it is most similar to the South American *S. aculeocrepitans*. It differs from the latter species mainly by longer spines, which tend to fuse together and produce shallow, wide pores.

Specimens examined. China. Yunnan: Xishuangbanna, Mandian Waterfalls, hardwood (?), 17.VIII.2005 Miettinen 10380.1* (H). Indonesia. Riau: Indragiri Hulu, Daerah Sungai Denalo, dead polypore, 2.VII.2004 Miettinen 8829 (H). Thailand. Phang Nga: Kurabari, Kura Andaman Private Beach, hardwood, 3.II.2015 Dunaev KUN1105* (H). Singapore. Yishun, Nee Soon pipeline, hardwood, 17.III.2020 Miettinen 23533* (SING, H).

***Sistotremastrum aculeocrepitans* Gruhn & Alvarado, Phytotaxa 379: 32, 2018.**—Fig. 10

Basidiocarps effused, compact, covering several cm, 0.1–0.2 mm thick. Hymenial surface cream-colored to pale ochraceous, odontoid; spines densely arranged, 3–4 per mm, up to 0.3 mm long, sharp-pointed, sometimes fusing together and then fimbriate. Margin white to pale cream-colored, adnate, producing abundant thin, short, white hyphal strands. Smooth areas between spines fertile.

Hyphal structure monomitic; hyphae clamped. Subicular hyphae interwoven, distinctly thick-walled, branched mostly at sharp angles, (3.4–) 3.8–5.7 (–6.0) μm in diam. ($n = 20/1$). Tramal hyphae distinctly thick-walled, densely arranged and

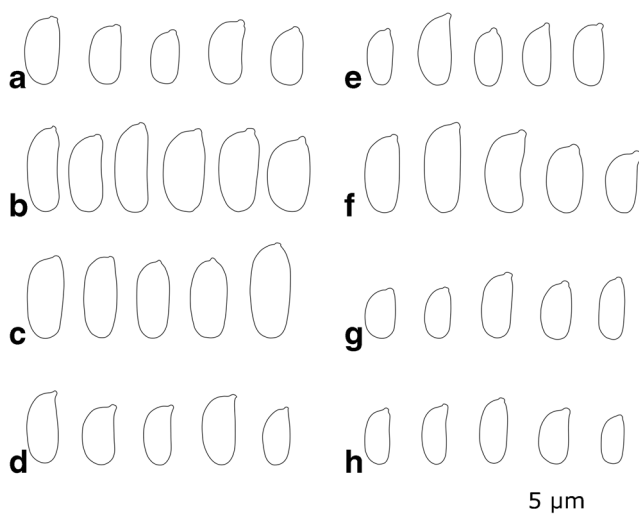


Fig. 10 Basidiospores of *Sertulicium* and *Sistotremastrum* spp. **a** *Sertulicium granuliferum* (Miettinen 14813.2). **b** *Sertulicium jacksonii* (three left—holotype, three right—Svantesson 699). **c** *S. niveocremaum* (holotype). **d** *Sistotremastrum aculeatum* (holotype). **e** *S. aculeocrepitans* (K.H. Larsson 16097). **f** *S. confusum* (holotype). **g** *S. denticulatum* (holotype). **h** *S. fibrillosum* (K.H. Larsson 16988)

partly glued together, (2.8–) 2.9–4.8 (–5.2) μm in diam. ($n = 20/1$), in subhymenium more or less vertically oriented, thin- or slightly thick-walled, short-celled and slightly inflated, (2.9–) 3.1–4.8 (–4.9) μm ($n = 20/1$). Hyphidia abundant, embedded or projecting up to 20 μm , simple, 2–3.5 μm in diam. Basidia suburniform, 4-spored, (7.7–) 8.4–13.8 (–14.2) \times (4.0–) 4.1–5.0 (–5.2) μm ($n = 30/2$). Basidioles clavate to bladder-shaped, 7–9 \times 4–5 μm . Basidiospores cylindrical to narrowly ovoid, straight, a few of the longest spores slightly lacrymoid, (4.0–) 4.1–5.6 (–5.8) \times (2.1–) 2.2–2.8 (–2.9) μm ($n = 60/2$), $L = 4.44$ –5.10, $W = 2.30$ –2.35, $Q = 1.93$ –2.19.

Remarks. The species was introduced based on specimens from Martinique and French Guiana (Gruhn et al. 2018). We collected it in Brazil, and these specimens are the source of the present description. Differences of *S. aculeocreptans* from *S. denticulatum* are treated under the latter species. Another similar-looking species from South America, *S. fibrillosum* Gruhn & Alvarado, has soft basidiocarps and densely arranged spines covering hyphal strands, and also considerably longer basidia. *Sistotremastrum aculeocreptans* is reported here as new to Brazil.

Specimens examined. Brazil. Paraíba: Areia, Reserva Mata do Pau-Ferro, angiosperm, 28.IV.2013 K.H. Larsson 16097* (URM, O). Pará: Belém, Museu Goeldi, Campus de Pesquisa, angiosperm, 25.XI.2013 K.H. Larsson 16478* (MG, O).

***Sistotremastrum confusum* K.H. Larss. & Spirin, sp. nov.**—Fig. 10

MB 833949

Holotype. Brazil. Pernambuco: Jaqueira, Reserva Particular do Patrimônio Natural Frei Caneca, Barragem das Moças, -8.720420° -35.843436°, on rotten angiosperm wood, 20.IV.2013 Larsson 16023* (URM, isotype—O).

Etymology. *Confusus* (Lat., adj.)—confusing.

Basidiocarps effused, soft, covering several cm, 0.04–0.07 mm thick. Hymenial surface whitish to cream-colored, smooth or irregularly tuberculate, accidentally cracking in old basidiocarps. Margin first arachnoid, with well-visible thin hyphal strands, later compact and rather clearly delimited.

Hyphal structure monomitic; hyphae clamped. Subicular hyphae interwoven to subparallel, slightly thick-walled, irregularly branched, (2.2–) 2.3–4.2 (–4.8) μm in diam. ($n = 40/2$). Subhymenial hyphae thin- or slightly thick-walled, rather loosely arranged, some short-celled and slightly inflated, (2.4–) 2.6–4.2 (–4.8) μm in diam. ($n = 40/2$). Hyphidia rarely present, embedded or only slightly projecting, simple, 3–3.5 μm in diam. Basidia clavate, 4-spored, gradually tapering to the basal part, (12.2–) 13.1–25.0 (–28.8) \times (4.3–) 4.4–6.2 (–6.3) μm ($n = 22/2$). Basidioles clavate to bladder-shaped, (7.3–) 7.4–11.8 (–13.0) \times (4.2–) 4.3–5.3 (–6.0) μm ($n = 20/2$). Basidiospores cylindrical to narrowly ellipsoid or narrowly ovoid, (5.3–) 5.7–7.6 (–7.9) \times 2.9–3.9 (–4.0) μm ($n = 60/2$), L

$= 6.54$ –6.57, $W = 3.16$ –3.32, $Q = 1.99$ –2.07.

Remarks. *Sistotremastrum confusum* is introduced here based on two sequenced collections from Brazil. Morphologically, it is most similar to *S. vigilans* distributed in Northern Hemisphere. The latter species has considerably larger basidia and basidiospores than those of *S. confusum*.

Specimens examined. Brazil. Pernambuco: Jaqueira, Reserva Particular do Patrimônio Natural Frei Caneca, Barragem das Moças, angiosperm, 20.IV.2013 K.H. Larsson 16004* (URM, O).

***Sistotremastrum denticulatum* Motato-Vásquez & Spirin, sp. nov.**—Fig. 10

MB 833950

Holotype. Brazil. São Paulo: Iguape, Juréia-Itatins Ecological Station, -24.417° -47.372°, fallen branch, 19.II.2017 Motato-Vásquez 894* (SP 467087, isotype—H 7009716).

Etymology. *Denticulatus* (Lat., adj.)—denticulate.

Basidiocarps effused, soft, waxy covering several cm, 0.05–0.1 mm thick. Hymenial surface cream-colored to pale ochraceous, odontoid; spines densely arranged, 5–6 per mm, up to 0.5 mm long, rather blunt-pointed, sometimes fusing together and then fimbriate. Margin white to pale cream-colored, adnate, arachnoid, producing abundant thin, short, white hyphal strands. Smooth areas between spines fertile.

Hyphal structure monomitic; hyphae clamped. Subicular hyphae interwoven to subparallel, distinctly thick-walled, branched mostly at sharp angles, (2.6–) 3.2–5.2 (–5.4) μm in diam. ($n = 20/1$). Tramal hyphae interwoven to subparallel, distinctly thick-walled, rather loosely arranged, (3.1–) 3.2–4.2 (–4.8) μm in diam. ($n = 20/1$), in subhymenium more or less vertically oriented, thin- or slightly thick-walled, mostly short-celled and slightly inflated, (2.2–) 2.3–3.8 (–4.1) μm ($n = 20/1$). Hyphidia abundant, mostly slightly projecting, simple, 2–3 μm in diam. Basidia suburniform to almost clavate, (1–) 2–4-spored, (6.7–) 7.2–12.2 (–12.3) \times (3.7–) 3.8–4.8 (–5.0) μm ($n = 20/1$). Basidioles clavate to subglobose, (4.8–) 5.2–7.2 (–7.6) \times (3.5–) 3.8–4.8 (–4.9) μm ($n = 20/1$). Basidiospores narrowly ellipsoid to broadly cylindrical, straight, a few of the longest spores somewhat fusiform, (3.9–) 4.0–5.3 (–5.4) \times (2.0–) 2.1–3.0 (–3.1) μm ($n = 30/1$), $L = 4.51$, $W = 2.48$, $Q = 1.83$.

Remarks. *Sistotremastrum denticulatum* is morphologically most similar to *S. aculeocreptans* but it has more densely arranged spines fusing together in characteristic fimbriate groups. Microscopically, these species are almost identical except for loosely arranged and not agglutinated hyphae in *S. denticulatum*. *Sistotremastrum fibrillosum* has spines not only on the hymenial surface but also on hyphal cords while *S. denticulatum* has sterile hyphal strands. The species is so far known only from the type locality in the southern part of Brazil.

Sistotremastrum fibrillosum Gruhn & Alvarado, Phytotaxa 379: 31, 2018—Fig. 10

Basidiocarps effused, soft, covering several cm, 0.05–0.1 mm thick. Hymenial surface cream-colored, odontoid; spines densely arranged, 6–9 per mm, up to 0.4 mm long, sharp-pointed, often fusing together in fascicles of 2–4, covering also hyphal strands. Margin white to pale cream-colored, floccose or radially fibrillose. Hyphal strands well developed, sometimes a few cm long and up to 0.5 mm thick.

Hyphal structure monomitic; hyphae clamped. Subicular hyphae mostly subparallel, distinctly thick-walled, branched mostly at sharp angles, (3.3–) 3.6–6.2 (–7.1) μm in diam. ($n = 20/1$); some hyphae encrusted by densely distributed subglobose resinous globules. Tramal hyphae with unevenly thickened walls, rather loosely arranged, subparallel to interwoven, (3.2–) 3.3–4.3 (–4.6) μm in diam. ($n = 20/1$), in subhymenium more or less vertically oriented, thin- or slightly thick-walled, sometimes short-celled and slightly inflated, (1.7–) 2.0–3.2 (–3.6) μm ($n = 20/1$). Hyphidia present, embedded or projecting up to 20 μm , simple, 1.5–2 μm in diam. Basidia suburniform, 4-spored, (9.4–) 10.3–19.2 (–20.8) \times (4.0–) 4.1–4.8 (–5.1) μm ($n = 20/1$). Basidioles clavate to bladder-shaped, 7–10 \times 3.5–4.5 μm . Basidiospores cylindrical to narrowly ellipsoid, straight, (4.0–) 4.2–5.2 (–5.6) \times (2.1–) 2.2–2.9 (–3.0) μm ($n = 20/1$), $L = 4.72$, $W = 2.53$, $Q = 1.88$, often glued together in large groups.

Remarks. The description above is based on a single specimen from Brazil. *Sistotremastrum fibrillosum* was originally introduced from four specimens collected in French Guiana (Gruhn et al. 2018). However, available DNA sequences show considerable variation, and this may indicate *S. fibrillosum* in the current sense is a collective species. More material is needed to find a more solid solution. Morphological differences of *S. fibrillosum* from *S. aculeocrepitans* detected in the same geographic region are treated under the latter species.

Specimens examined. Brazil. Pará: Melgaço, Caxiuanã, ICMBIO headquarter, angiosperm, 13.III.2015 *K.H. Larsson* 16988* (MG 213771, O).

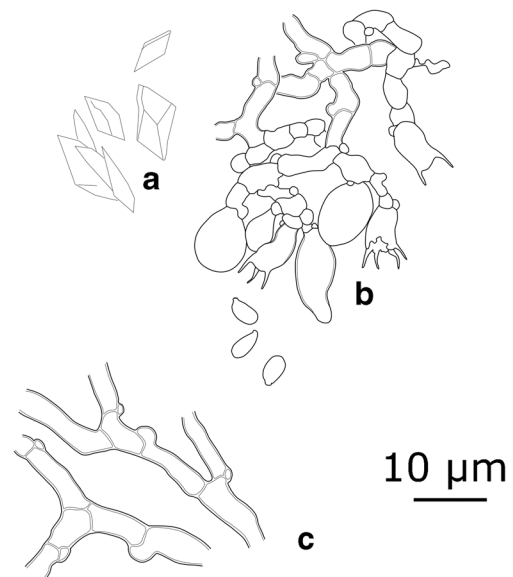


Fig. 12 Microscopic structures of *S. geminum* (holotype). **a** Crystals from subhymenial hyphae. **b** Subhymenial hyphae and hymenial cells. **c** Subicular hyphae

Sistotremastrum geminum Miettinen & Spirin, **sp. nov.**—Figs. 11 and 12

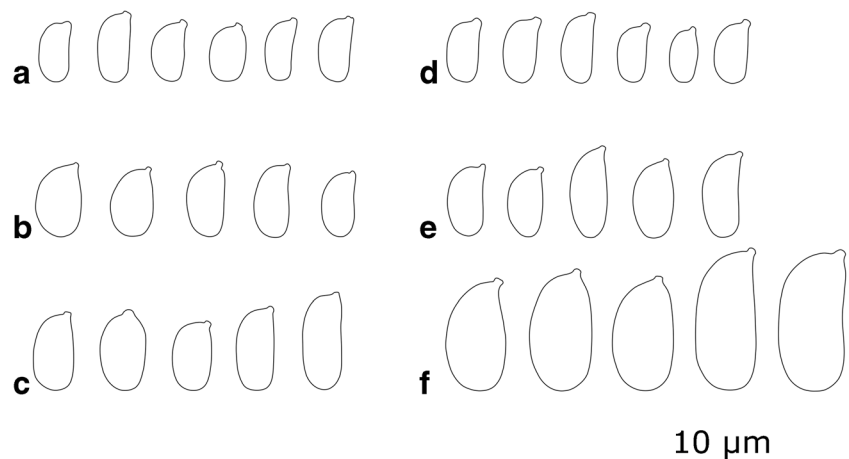
MB 833951

Holotype. Indonesia. Papua Barat: Manokwari Reg., Saukorem, Tamuk, -0.5832° 133.14845° , hilly lowland primary rainforest, on an uprooted *Intsia bijuga* (150 cm in diameter, decay stage 1–4/5), 30.X.2010 *Miettinen* 14333.2* (MAN, isotype—H7009307).

Etymology. *Geminus* (Lat., adj.)—similar.

Basidiocarps effused, soft-floccose, covering several cm, 0.07–0.1 mm thick. Hymenial surface pale cream-colored, grandinoid, with irregularly arranged, obtuse warts 7–8 per mm. Margin pale cream-colored, floccose, partly detaching. Hyphal strands abundant in the underlying substrate, white and richly branched. Smooth areas between warts fertile.

Fig. 11 Basidiospores of *Sistotremastrum* spp. **a** *S. geminum* (holotype). **b** *S. induratum* (holotype). **c** *S. mendax* (three left—holotype, three right—Spirin 10060). **d** *S. rigidum* (holotype). **e** *S. suecicum* (holotype). **f** *S. vigilans* (three left—Spirin 10097, two right—holotype)



Hyphal structure monomitic; hyphae clamped. Subicular hyphae interwoven to subparallel, slightly thick-walled, (2.3–) 2.8–4.8 (–5.2) μm in diam. ($n = 20/1$). Tramal hyphae thin-walled, rather loosely arranged, interwoven, as a rule short-celled and more or less distinctly inflated, (3.0–) 3.1–4.8 (–6.2) μm in diam. ($n = 20/1$). Subhymenial hyphae thin-walled, easily collapsing, often short-celled and slightly inflated or twisted, (2.2–) 2.3–3.4 (–3.7) μm in diam. ($n = 20/1$). Hyphal ends at spine tips thin-walled, short-celled, with occasional short side branches, 3–5 μm in diam., usually encrusted by solitary rhomboid crystals or solid crystalline shields. Cystidia unevenly distributed, thin-walled, varying from obtuse-clavate to subglobose, mostly enclosed, $11\text{--}22 \times 6.5\text{--}9.5$ μm . Hyphidia present, embedded or slightly projecting, simple, 2–3 μm in diam. at the apex, sometimes swollen at the base up to 4 μm in diam. Basidia suburniform, 4-spored, (7.8–) 8.0–11.2 (–11.8) \times (3.6–) 3.7–4.4 (–4.6) μm ($n = 20/1$), senescent basidia slightly thick-walled at the basal part. Basidioles bladder-shaped to subglobose, (4.8–) 4.9–6.8 (–7.2) \times (3.6–) 3.9–4.7 (–5.3) μm ($n = 20/1$). Basidiospores cylindrical to narrowly ellipsoid, straight to slightly curved, (3.8–) 3.9–5.2 (–5.5) \times (1.9–) 2.0–2.6 (–2.9) μm ($n = 30/1$), $L = 4.28$, $W = 2.24$, $Q = 1.92$.

Remarks. Hymenophore with short warts (as opposed to long spines) and thin-walled, loosely arranged tramal hyphae differentiate *S. geminum* from other four-sterigmatic representatives of the genus treated here. The species is so far known only from the type locality in New Guinea.

***Sistotremastrum induratum* Spirin, sp. nov.—Fig. 11**

MB 833952

Holotype. USA. Washington: Pend Oreille Co., Muskegon Lake, 48.7923° –117.044°, *Abies grandis*, 16.X.2014 Spirin 8598* (H 7009717).

Etymology. *Induratus* (Lat., adj.)—hardening.

Basidiocarps effused, initially pruinose, later compact, covering several cm, 0.03–0.1 mm thick. Hymenial surface whitish to cream-colored, smooth or porulose. Margin first pruinose, later compact and rather sharply delimited. Hyphal strands present in subiculum of vigorously growing specimens, tough and well-differentiated.

Hyphal structure monomitic; hyphae clamped. Hyphae in strands densely arranged and partly agglutinated, more or less parallel, unevenly thick-walled, 5–6 μm in diam. Subicular hyphae interwoven to subparallel, unevenly thick-walled, branched mostly at right angles, (2.7–) 2.8–4.5 (–4.7) μm in diam. ($n = 20/1$). Subhymenial hyphae thin- or slightly thick-walled, densely arranged, some short-celled and slightly inflated, (2.8–) 3.0–4.8 (–5.0) μm in diam. ($n = 40/2$). Hyphidia occasionally present, embedded, simple, 3–5 μm in diam. Basidia clavate, 4–6-spored, gradually tapering to the basal part, (13.6–) 14.0–22.8 (–24.0) \times (5.1–) 5.2–6.3 (–6.8) μm ($n = 20/1$). Basidioles clavate to bladder-shaped, (9.8–)

10.2–15.9 (–16.0) \times (3.8–) 3.9–5.3 (–5.7) μm ($n = 20/1$). Basidiospores broadly cylindrical to narrowly ovoid, rarely ellipsoid, (4.0–) 4.1–5.3 (–5.8) \times (2.5–) 2.6–3.1 (–3.2) μm ($n = 60/2$), $L = 4.37\text{--}4.73$, $W = 2.79\text{--}2.86$, $Q = 1.53\text{--}1.70$.

Remarks. *Sistotremastrum induratum* has so far been detected in two localities in the North American North-West. It is most similar to the European *S. mendax* but it has on average shorter basidia and basidiospores, as well as stable differences in ITS sequences. Distribution areas of the two species seem not to overlap.

Specimens examined. USA. Washington: Jefferson Co., Hoh River, *Picea sitchensis*, 20.X.2014 Spirin 8804* (H).

***Sistotremastrum mendax* Spirin & Volobuev, sp. nov.—Figs. 5 and 11**

MB 833953

Holotype. Norway. Telemark: Nome, Mørkvasslia Nat. Res., 59.288644° 8.865084°, rotten log of *Picea abies*, 23.IX.2003 K.H. Larsson 12022* (GB).

Etymology. *Mendax* (Lat., adj.)—deceptive.

Basidiocarps effused, initially pruinose, later compact, covering several cm, 0.05–0.2 mm thick. Hymenial surface whitish grey to cream-colored or pale ochraceous, smooth or porulose, accidentally cracking only in very old basidiocarps. Margin first pruinose, later compact and rather sharply delimited. Hyphal strands present in subiculum of vigorously growing specimens, normally detectable by the naked eye.

Hyphal structure monomitic; hyphae clamped. Hyphae in strands densely arranged, parallel, slightly thick-walled, 1.5–2.5 μm in diam. Subicular hyphae interwoven to subparallel, slightly thick-walled, irregularly branched, (3.1–) 3.7–6.2 (–6.3) μm in diam. ($n = 20/1$). Subhymenial hyphae thin- or slightly thick-walled, densely arranged, some short-celled and slightly inflated, (2.2–) 2.3–4.2 (–4.3) μm in diam. ($n = 50/3$). Hyphidia rarely present, embedded, simple, 3–4 μm in diam. Basidia clavate, 4–6-spored, gradually tapering to the basal part, (11.4–) 11.7–27.2 (–27.3) \times (4.6–) 4.8–6.6 (–7.2) μm ($n = 50/3$). Basidioles clavate to bladder-shaped, (7.2–) 8.2–13.8 (–15.3) \times (4.2–) 4.3–6.2 (–6.5) μm ($n = 20/2$). Basidiospores narrowly ellipsoid to broadly cylindrical, some narrowly ovoid, (4.0–) 4.2–7.0 (–7.2) \times (2.4–) 2.5–3.5 (–3.6) μm ($n = 180/6$), $L = 4.75\text{--}5.71$, $W = 2.76\text{--}3.09$, $Q = 1.71\text{--}1.95$.

Remarks. The species is introduced here as a sibling of *S. suecicum* occurring on wood of *Picea* in Europe. It differs from *S. suecicum* s. str. in having broader basidiospores and much better developed and easily observable hyphal strands in subiculum. Moreover, mature basidiocarps of *S. suecicum* are cracking in a characteristic way while they mostly stay continuous in *S. mendax* (Fig. 2). The host preference alone is not enough to recognize these species because *S. suecicum* sometimes occurs on spruce, as well as other coniferous trees. The

North American *S. induratum* has on average shorter basidia and basidiospores than *S. mendax* (Table 2).

According to our observations, *S. mendax* prefers old-growth spruce-dominated forests subjected to minimal (if any) logging activity. Further ecological studies are needed to confirm these observations.

Specimens examined. Czech Republic. South Bohemia Reg.: Boubín Nat. Res., *P. abies*, 29.X.2011 Vlasák 1110/14.1 (H). Finland. Etelä-Karjala: Miehikkälä, Kaitai, *P. abies*, 3.VIII.1976 Fagerström (H). Satakunta: Lamppi, *P. abies*, 31.VIII.1939 Laurila (H). Norway. Sør-Trøndelag: Neadalen, Tydal, *P. abies*, 24.IX.2011 J. Nordén 9579* (O). Poland. Podlasie: Hajnówka, Białowieża NP, *P. abies*, 23.IX.2019 Miettinen 22476* (H). Russia. Karelia: Medvezhiegorsk Dist., Krivozero, *P. abies*, 7.VI.1942 Laurila 3251a, 3258 (H). Leningrad Reg.: Boksitogorsk Dist., Chagoda, *P. abies*, 9.V.2018 Spirin 11887 (H), Kolp', *P. abies*, 8.V.2016 Spirin 10057, 10060 (H), 9.V.2016 Spirin 10086 (H); Podporozhie Dist., Oksboloto, *P. abies*, 27.V.2017 Spirin 11223 (H), Vazhinka, *P. abies*, 21.V.2016 Spirin 10107 (H), 4.VI.2016 Spirin 10151 (H), 21.VI.2016 Spirin 10226 (H), 26.V.2017 Spirin 11208 (H), Miettinen 20946* (H), 14–15.X.2017 Spirin 11699, 11738 (H). Sweden. Värmland: Torsby, *P. abies*, 5.X.2010 J. Nordén 7350 (O).

***Sistotremastrum rigidum* Motato-Vásquez & Spirin, sp. nov.**—Fig. 11

MB 833954

Holotype. Brazil. São Paulo: Trilha do Mirante, Serra do Mar State Park, -23.983° -46.7467°, fallen log, 17.XI.2016 Motato-Vásquez 833* (SP 466010, isotype—H).

Etymology. *Rigidus* (Lat., adj.)—rigid, tough.

Basidiocarps effused, crustaceous, tough, covering several cm, 0.05–0.08 mm thick. Hymenial surface almost white to cream-colored, smooth. Margin white, indistinctly fimbriate, adnate, occasionally with short hyphal strands.

Hyphal structure monomitic; hyphae clamped. Subicular hyphae tightly glued together, interwoven to subparallel, thick-walled, hardly visible, 2–4 µm in diam. Subhymenial hyphae slightly to distinctly thick-walled, densely arranged and partly glued together, some short-celled and slightly inflated, (2.0–) 2.1–3.0 (–3.2) µm in diam. (n = 20/1). Hyphidia rarely present, mostly embedded, simple, 2–2.5 µm in diam. Basidia suburniform to almost clavate, 4-spored, (7.7–) 9.2–14.3 (–16.2) × (4.6–) 4.8–6.6 (–7.2) µm (n = 20/1), senescent basidia slightly thick-walled at the basal part. Basidioles rare, subglobose, 5–6 × 4–5 µm. Basidiospores cylindrical to broadly cylindrical, (3.8–) 4.0–5.1 (–5.2) × 2.0–2.5 (–2.7) µm (n = 30/1), L = 4.44, W = 2.22, Q = 2.01.

Remarks. Basidiocarps of *S. rigidum* are exceptionally tough due to densely arranged and partly agglutinated hyphae,

making the structure resemble species of *Phlebia* s.l. However, none of the latter ones forms hyphal strands or has suburniform basidia with slightly but distinctly thickened walls. Another four-sterigmatic species with smooth hymenophore, *S. confusum*, occurs in Brazil. However, it differs from *S. rigidum* in having soft basidiocarps and larger basidia and basidiospores.

***Sistotremastrum suecicum* Litsch. ex J. Erikss., Symb. Bot. Upsalienses 16 (1): 62, 1958—Figs. 5 and 11**

Holotype. Sweden. Upland: Uppsala, Bondkyrka, “on the under-side of coniferous fencing-material, lying in a wet spot” [*Pinus sylvestris*—identified by Pirkko Harju, H], 4.I.1933 Lundell (Fungi Exsiccati Suecici #464) (S F204406, studied).

Basidiocarps effused, initially pruinose, later waxy, covering several cm, 0.05–0.2 mm thick. Hymenial surface whitish to cream-colored or pale ochraceous, smooth or porulose, in older basidiocarps often cracking into small irregular patches. Margin first pruinose, later compact and rather sharply delimited. Hyphal strands present in basal layer of vigorously growing specimens.

Hyphal structure monomitic; hyphae clamped. Hyphae in strands densely arranged, parallel, slightly thick-walled, 1.5–2.5 µm in diam. Subicular hyphae interwoven to subparallel, slightly thick-walled, branched mostly at right angles, (3.8–) 3.9–5.8 (–6.2) µm in diam. (n = 40/2). Subhymenial hyphae thin- or slightly thick-walled, densely and randomly arranged, in subhymenium more or less vertically oriented, some short-celled and slightly inflated, (2.2–) 2.3–4.1 (–4.2) µm in diam. (n = 100/5). Hyphidia occasionally present, embedded or projecting (especially in young specimens) up to 15 µm above hymenial layer, simple, 2–4 µm in diam. Basidia clavate, 4–6-spored, gradually tapering to the basal part, (10.6–) 12.8–26.7 (–26.8) × (4.1–) 4.2–6.2 (–6.3) µm (n = 80/5). Basidioles clavate to bladder-shaped, (8.3–) 8.6–14.2 (–15.1) × (3.3–) 3.8–5.2 (–5.8) µm (n = 20/1). Basidiospores cylindrical to narrowly ovoid, (4.1–) 4.2–6.2 (–6.3) × (2.1–) 2.2–3.1 (–3.2) µm (n = 180/6), L = 4.56–5.55, W = 2.42–2.80, Q = 1.83–2.06.

Remarks. *Sistotremastrum suecicum* was described from Sweden (Eriksson 1958) and has so far been treated in a collective sense, i.e., the name has been used for all collections from coniferous hosts. According to our results, *S. suecicum* occurs almost exclusively on *Pinus* spp., like the so-called kelo trees in North Europe (Niemelä et al. 2002). It is considered an indicator species of old pine forests of high conservation value in Finland (Niemelä 2016). Most specimens from other gymnosperm hosts belong either to *S. mendax* (in Europe) or to *S. induratum* (in the North American North-West). Differences of *S. suecicum* from the latter two species are discussed under *S. mendax*. The species forms basidiocarps throughout the year.

An ITS sequence of *S. suecicum* UC2022903 found on pine wood in California (GenBank KP814148) is clearly different from the rest of the pine-dwelling collections and seemingly representing a species of its own. We leave this problem until more material from the US Pacific will be available. Future studies should elucidate if *S. suecicum* s. str. is also present in this geographic area or if its distribution in North America is limited to the east of the Rocky Mountains.

Specimens examined. Canada. British Columbia: Fraser—Fort George Reg. Dist., Jackman Flats Prov. Park, *Pinus contorta*, 25.VII.2015 *Spirin* 8932* (H). Nova Scotia: Kentville, Ravine, *Pinus resinosa*, 8.VIII.1954 *Harrison* (H ex DAOM 52237). Finland. Varsinais-Suomi: Parainen, Kuggö, *P. sylvestris*, 24.X.2009 *Kunttu* 5959* (H). Pohjois-Savo: Heinävesi, Vihtarinniemi, *P. sylvestris*, 2.X.2006 *Miettinen* 11044.1* (H). Inarin Lappi: Inari, Piekanaavaara, *P. sylvestris*, 28.VIII.2008 *Miettinen* 13310.1 (H). Russia. Arkhangelsk Reg.: Krasnoborsk Dist., *P. sylvestris*, 5.VIII.2013 *Kotkova* (LE 295792*). Krasnoyarsk Reg.: Turukhansk Dist., Bor, *P. sylvestris*, 24.VIII.2013 *Kotiranta* 26451 (H). Leningrad Reg.: Boksitogorsk Dist., Chagoda, *P. sylvestris*, 10.V.2018 *Spirin* 11888, 11890 (H). Shidrozero, *P. sylvestris*, 14.VIII.2014 *Spirin* 6974 (H); Podporozhie Dist., Kurba, *P. sylvestris*, 19.IX.2009 *Spirin* 3236 (H), Vazhinka, *P. abies*, 14.X.2017 *Spirin* 11697 (H). Nizhny Novgorod Reg.: Arzamas Dist., Pustyn', *P. sylvestris*, 13.VIII.2015 *Spirin* 9227, 9233 (H). Sweden. Jämtland: Härjedalen, Glumtjärnarna, *P. sylvestris*, 14.VIII.2011 *Miettinen* 14550.1* (H). Ångermanland: Resele, Vignäsbodarna, *P. abies* (?), 28.VIII.2002 *K.H. Larsson* 11849* (GB). Jämtland: Härjedalen, Ransjön, *P. sylvestris*, 14.VIII.2011 *Miettinen* 14548 (H). USA. Massachusetts: Worcester Co., Holden, fallen branch, 26.IX.2011 *Miettinen* 14829* (H), *Tsuga canadensis*, 21.IV.2013 *Miettinen* 16061* (H), *Tsuga/Pinus* sp., 27.VII.2013 *Miettinen* 16618* (H).

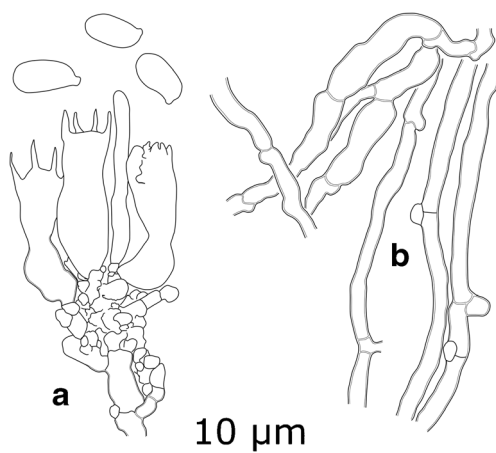


Fig. 13 Microscopic structures of *S. vigilans* (holotype). **a** Subhymenial hyphae and hymenial cells. **b** Subicular hyphae

Sistotremastrum vigilans K.H. Larss. & Spirin, **sp. nov.**—Figs. 11 and 13

MB 833955

Holotype. Norway. Vestfold: Stokke, Melsomvik, 59.22541° 10.35202°, rotten log of *Picea abies*, 30.IV.2011 *Fonneland* 2011-78* (O).

Etymology. *Vigilans* (Lat., adj.)—vigilant.

Basidiocarps effused, compact, covering several cm, 0.05–0.2 mm thick. Hymenial surface white to cream-colored or pale ochraceous, first smooth, in older basidiocarps irregularly cracking and unevenly tuberculate. Margin first pruinose, later compact and sharply delimited. Hyphal strands often present, well-developed, white, occurring in subiculum and sometimes in the underlying substrate.

Hyphal structure monomitic; hyphae clamped. Subicular hyphae in parallel bundles or interwoven, with unevenly thickened walls, randomly branched, some tortuous, with occasional oil-drops, (3.1–) 3.2–5.6 (–5.7) μm in diam. (n = 60/3), in older parts with bubble-like inflations 7–10 μm in diam. Subhymenial hyphae thin- or slightly thick-walled, densely arranged, some short-celled and slightly inflated, (2.0–) 2.2–4.2 (–5.0) μm in diam. (n = 60/3). Hyphidia occasionally present, embedded or projecting up to 20 μm, simple or with accidental side branches, 1.5–5 μm in diam. Basidia clavate to utriform, 4-spored, with widened or tapering bases, (11.3–) 11.4–34.1 (–38.2) × (5.3–) 5.4–8.0 (–8.1) μm (n = 50/3). Basidiospores narrowly ellipsoid to long cylindrical, straight or slightly curved, a few lacrymoid or sigmoid, (6.1–) 6.3–11.2 (–11.4) × (3.0–) 3.1–5.4 (–6.2) μm (n = 150/5), L = 7.43–9.95, W = 3.64–4.69, Q = 1.81–2.22.

Remarks. *Sistotremastrum vigilans* is the only four-sterigmatic species of the genus so far detected in temperate and boreal forests of the Northern Hemisphere. It occurs on coniferous hosts, either late in the season or in spring, and this might have been a reason for overlooking it. The species is rare and tends to occur in rather old forests with large-diameter dead wood.

Sistotremastrum vigilans has the largest basidiospores in the genus and thus it can be mistaken for a *Hyphoderma* s. l. species. Basidia of highly variable size, basidiospores with homogeneous contents (not guttulate as in *Hyphoderma*) and slightly thicker walls, as well as occasionally inflated subicular hyphae are the most reliable traits for a morphological identification of this species. The well-developed hyphal strands (if present) are another clue to recognizing *S. vigilans*.

Specimens examined. Canada. Québec: Outaouais, Gatineau Nat. Park, *Abies balsamea*, 14.X.1967 *Eriksson* 9106 (GB, K), conifer, 15.X.1967 *Eriksson* 9140, 9270, 9279 (GB, K), *A. balsamea*, 22.X.1967 *Eriksson* 9281 (GB, K). Finland. Uusimaa: Helsinki, Laajasalo, *P. abies*, 25.XI.1934 *Häyrén* (H), Haltiala, *P. abies*, 20.XII.2020 *Miettinen* 24513 (H). Satakunta: Vuorijärvi, *P. abies*, 6.IX.1937 *Laurila* (H). Norway. Vestfold: Stokke,

Melsomvik, *P. abies*, 30.IV.2011 *Fonneland 2011-77* (O). Russia. Leningrad Reg.: Podporozhie Dist., Vazhinka, *P. abies*, 21.V.2016 *Spirin 10097** (H), 22.V.2016 *Spirin 10135* (H). USA. Washington: Jefferson Co., Hoh River, *Tsuga heterophylla*, 20.X.2014 *Spirin 8778** (H).

Discussion

In this paper, we present the most comprehensive study of *Sistotremastrum* to date. *Sistotremastrum* sensu lato now holds 18 species, which means that species number more than doubled. The original concept of *Sistotremastrum* (Eriksson 1958) contained only two species, *S. suecicum* and *S. niveocreumum*. However, with the much richer sampling available to us, the phylogenetic analyses prompted us to segregate the species around *S. niveocreumum* into the new genus *Sertulicium*.

Larsson (2007) showed that *Sistotremastrum suecicum* and *S. niveocreumum* belong in the Trechisporales forming a strongly supported sister clade to the rest of the order. Larsson (2007) named this clade the *Sistotremastrum* family while for the sister clade the name *Hydnodontaceae* is available. The same strongly supported topology was recovered by Gruhn et al. (2018) in a paper where two new *Sistotremastrum* species were introduced. In the present work, we were not able to generate high support values for the node connecting *Sistotremastrum* and *Sertulicium* (Fig. 1). The reason for this is unclear but probably depends on the generally high diversity of the nuclear ribosomal markers in Trechisporales. We think it is likely that the concept of *Sistotremastrum* family as defined by Larsson will survive, but the question can only be solved with sequences from an expanded set of genes.

Three species, *Sistotremastrum denticulatum*, *S. geminum*, and *S. rigidum*, are described based on single specimens. While we normally would avoid doing this, we found it justified in these cases. These three species are all well separated in ITS alignments and differs by 3.6%, 4.6%, and 9.0%, respectively, from their phylogenetically closest relatives (Fig. 1). Furthermore, all the close relatives of these single-specimen species are represented by multiple ITS sequences, increasing confidence that such big ITS differences represent inter- and not infraspecific variation. All these species are also morphologically clearly separable from their closest relatives. Lastly, we used distinct distribution areas as a criterion. Distribution areas of *S. denticulatum* and *S. rigidum* do not overlap with their closest relatives. Difference in distribution areas is not as clear in the case of *S. geminum* and its closest relative *S. aculeatum*, which are both found in South-East Asia. However, the latter has been found so far only west of Wallace's line (Sumatra and Asian mainland), while *S. geminum* east of it, in Australasia (New Guinea).

Other specimens examined

Brevicellicium exile. USA. Washington: Clark Co., Gifford Pinchot Nat. Forest, *T. heterophylla*, 11.X.2014 *Spirin 8370** (H).

B. olivascens. Russia. Nizhny Novgorod Reg.: Bogorodsk Dist., Krastelikha, *Q. robur*, 5.IX.2011 *Spirin 4446** (H). Sweden. Bohuslän: Restenäs, Ulvesund, decaying angiosperm branch, 3.X.1998, *Larsson 8571** (GB).

B. viridulum. Russia. Sakhalin: Uglegorsk Dist., Telnovskoe, *Alnus hirsuta*, 1.IX.2007 *Kotiranta 29271** (H).

Luellia cystidiata. Denmark. Jylland: Varde, Bordrup Klitplantage, *P. abies*, 6.X.2009 *Petersen & Læssøe 13875* (GB).

Paullicorticium allantosporum. Norway. Hedmark: Løten, Gitvola, *P. abies*, 26.IX.2018 *Spirin 12413* (H).

Pteridomyces galzinii. Italy. Tuscany: Arezzo, Camaldoli Nat. Res., *Polystichum aculeatum*, 17.XI.2010 *Bernicchia 8122** (GB).

Sertulicium aff. *granuliferum*. Kenya. Eastern Prov.: Meru, Chigora, decayed wood, 31.I.1973 *Ryvarden 3593* (O); Nyeri, Regati, decayed wood, 2.II.1973 *Ryvarden 9777* (O), *Ryvarden 9888* (O). Tanzania. Tanga Prov.: Tanga Dist., Amani, decayed wood, 18.II.1973 *Ryvarden 10575* (O, H).

Sertulicium sp. 1. Russia. Khabarovsk Reg.: Khabarovsk Dist., Ilga, *Corylus mandshurica*, 11.VIII.2012 *Spirin 5158** (H).

Sertulicium sp. 2. Sweden. Bohuslän: Säve, *P. sylvestris* (bark), 4.III.1973 *Arvidsson* (GB).

Suillosporium cystidiatum. Russia. Khabarovsk Reg.: Solnechnyi Dist., Igdomi, *Picea ajanensis*, 5.VIII.2011 *Spirin 3830** (H).

Acknowledgements Greet Van Autgaerden (Antwerpen, Belgium), Eugeny Dunaev (Young Naturalist Club of the Zoological Museum, Moscow University, Russia), and Alexander Ordynets (Kassel, Germany) kindly provided us with valuable fungal collections. Serena Lee organized and participated in field work in Singapore. Pirkko Harju (Helsinki, Finland) identified host trees of some important specimens.

Author contribution All authors contributed to the study conception and design. Material preparation, data collection, and analysis were initially performed by Viacheslav Spirin, Sergey Volobuev, Ilya Viner, Otto Miettinen, and Karl-Henrik Larsson. The authors Josef Vlasák, Nathan Schoutteten, Viviana Motato-Vásquez, Heikki Kotiranta, and Hemawati provided further important material and/or sequences. The first draft of the manuscript was written by Viacheslav Spirin and all authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.

Funding Open access funding provided by University of Helsinki including Helsinki University Central Hospital. Komarov Botanical Institute RAS (institutional research project “Biodiversity, ecology, structural and functional features of fungi and fungus-like protists”, AAAA-A19-119020890079-6, with equipment of The Core Facilities Center “Cell and Molecular Technologies in Plant Science”) (St. Petersburg, Russia) (the

author SV); the grant of the President of the Russian Federation (project MK-3216.2019.11) (the author SV); Societas pro Fauna et Flora Fennica (the author IV); Academy of Sciences of the Czech Republic (project number RVO: 60077344) (the author JV); Flemish Research Council (FWO) (grant number 11E0420N) (the author NS); Coordenação de Aperfeiçoamento Pessoal de Nível Superior – CAPES (Brazil) (the author VMV); The Rufford Small Grants Foundation (the author VMV).

Data availability DNA sequences used in the present study are available in GenBank. Alignments were deposited in TreeBASE. Fungal specimens are stored in public herbaria (as indicated under “Specimens examined”).

Declarations

Ethics approval Not applicable

Conflict of interest The authors declare no competing interests.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Benson DA, Cavanaugh M, Clark K, Karsch-Mizrachi I, Ostell J, Pruitt KD, Sayers EW (2018) GenBank. *Nucleic Acids Res* 46(D1):D41–D47. <https://doi.org/10.1093/nar/gkx1094>
- Boidin J, Gilles G (1990) Corticiés s.l. intéressantes ou nouveaux pour la France (Basidiomycotina). *Bull Soc Mycol France* 106:135–167
- Boidin J, Gilles G (1994) Contribution à la connaissance des Corticiés à basides unifornes ou subunifornes (Basidiomycotina). *Bull Soc Mycol France* 110:185–229
- Eriksson J (1958) Studies in the Heterobasidiomycetes and Homobasidiomycetes – Aphyllophorales of Muddus National Park in North Sweden. *Symb Bot Upsal* 16:1–172
- Eriksson J, Ryvarden L (1975) The Corticiaceae of North Europe. 3. *Fungiflora*, Oslo
- Eriksson J, Hjortstam K, Ryvarden L (1978) The Corticiaceae of North Europe. 5. *Fungiflora*, Oslo
- Eriksson J, Hjortstam K, Ryvarden L (1984) The Corticiaceae of North Europe. 7. *Fungiflora*, Oslo
- Gardes M, Bruns TD (1993) ITS primers with enhanced specificity for basidiomycetes application to the identification of mycorrhizae and rusts. *Mol Ecol* 2:132–118. <https://doi.org/10.1111/j.1365-294x.1993.tb00005.x>
- Gouy M, Guindon S, Gascuel O (2010) SeaView version 4: a multiplatform graphical user interface for sequence alignment and phylogenetic tree building. *Mol Biol Evol* 27(2):221–224. <https://doi.org/10.1093/molbev/msp259>
- Gruhn G, Alvarado P, Hallenberg N, Roy M, Courtecuisse R (2018) Contribution to the taxonomy of *Sistotremastrum* (Trechisporales, Basidiomycota) and the description of two new species, *S. fibrillosum* and *S. aculeocephitans*. *Phytotaxa* 379:27–38
- Hallenberg N (1978) Wood-fungi (Corticiaceae, Coniophoraceae, Lachnocladiaceae, Thelephoraceae) in North Iran. I. *Iranian J Plant Pat* 14:38–87
- Haynes KA, Westerneng TJ, Fell JW, Moens W (1995) Rapid detection and identification of pathogenic fungi by polymerase chain reaction amplification of large subunit ribosomal DNA. *J Med Vet Mycol* 33(5):319–325. <https://doi.org/10.1080/02681219580000641>
- Huffman JA, Prenni AJ, DeMott PJ, Pöhlker C, Mason RH, Robinson NH, Fröhlich-Nowoisky J, Tobo Y, Després VR, Garcia E, Gochis DJ, Harris E, Müller-Germann I, Ruzene I, Schmer B, Sinha B, Day DA, Andreae MO, Jimenez JL, Gallagher M, Kreidenweis SM, Bertram AK, Pöschl U (2013) High concentration of biological aerosol particles and ice nuclei during and after rain. *Atmos Chem Phys* 13:6151–6164. <https://doi.org/10.5194/acp-13-6151-2013>
- Jülich W (1982) Notes on some Basidiomycetes (Aphylophorales and Heterobasidiomycetes). *Persoonia* 11(4):421–428
- Katoh K, Rozewicki J, Yamada KD (2017) MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. *Brief Bioinform*: bbb108. <https://doi.org/10.1093/bib/bbx108>
- Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, Thierer T, Ashton B, Meintjes P, Drummond A (2012) Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* 28(12):1647–1649. <https://doi.org/10.1093/bioinformatics/bts199>
- Larsson KH (1992) The genus *Trechispora*. University of Göteborg, Göteborg
- Larsson KH (2001) The position of *Poria mucida* inferred from nuclear ribosomal DNA sequences. *Harvard Papers Bot* 6:131–138
- Larsson KH (2007) Re-thinking the classification of corticioid fungi. *Mycol Res* 111:1040–1063
- Larsson KH, Larsson E, Køljalg U (2004) High phylogenetic diversity among corticioid homobasidiomycetes. *Mycol Res* 108:983–1002
- Liu SL, Ma HX, He SH, Dai YC (2019) Four new corticioid species in Trechisporales (Basidiomycota) from East Asia and notes on phylogeny of the order. *Mycologia* 111:97–113
- Menkis A, Allmer J, Vasilaiuskas R, Lygis V, Stenlid J, Finlay R (2004) Ecology and molecular characterization of dark septate fungi from roots, living stems, coarse and fine woody debris. *Mycol Res* 108: 965–973
- Miettinen O, Vlasák J, Rivoire B, Spirin V (2018) *Postia caesia* complex (Polyporales, Basidiomycota) in temperate Northern Hemisphere. *Fungal Syst Evol* 1:101–129
- Milne I, Lindner D, Bayer M, Husmeier D, McGuire G, Marshall DF, Wright F (2008) TOPALi v2: a rich graphical interface for evolutionary analyses of multiple alignments on HPC clusters and multi-core desktops. *Bioinformatics* 25(1):126–127. <https://doi.org/10.1093/bioinformatics/btn575>
- Nagy LG, Riley R, Tritt A, Adam C, Daum C, Floudas D, Sun H, Yadav JS, Pangilinan J, Larsson K-H, Matsuura K, Barry K, Labutti K, Kuo R, Ohm RA, Bhattacharya SS, Shirouzu T, Yoshinaga Y, Martin FM, Grigoriev IV, Hibbett DS (2016) Comparative genomics of early-diverging mushroom-forming fungi provides insights into the origins of lignocellulose decay capabilities. *Mol Biol Evol* 33(4):959–970. <https://doi.org/10.1093/molbev/msv337>
- Niemelä T (2016) Suomen käävät. *Norlinia* 31:1–430
- Niemelä T, Wallenius T, Kotiranta H (2002) The kelo tree, a vanishing substrate of specified wood-inhabiting fungi. *Polish Botanical Journal* 47:91–101
- Nilsson RH, Larsson K-H, Taylor AFS, Bengtsson-Palme J, Jeppesen TS, Schigel D, Kennedy P, Picard K, Glöckner FO, Tedersoo L, Saar I, Køljalg U, Abarenkov K (2018) The UNITE database for molecular identification of fungi: handling dark taxa and parallel taxonomic

- classifications. *Nucleic Acids Res* 47(D1):D259–D264. <https://doi.org/10.1093/nar/gky1022>
- Oberwinkler F (1965) Primitive Basidiomyceten. Revision einiger Formenkreise von Basidienpilzen mit plastischer Basidie. *Sydowia* 19:1–72
- Ronquist F, Teslenko M, Van Der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard M, Huelsenbeck JP (2012) MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Syst Biol* 61:539–542. <https://doi.org/10.1093/sysbio/sys029>
- Stamatakis A (2006) Raxml-vi-hpc: maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics* 22:2688–2690. <https://doi.org/10.1093/bioinformatics/btl446>
- Tamura K, Stecher G, Peterson D, Filipowski A, Kumar S (2013) MEGA6: molecular evolutionary genetics analysis version 6.0. *Mol Biol Evol* 30:2725–2729. <https://doi.org/10.1093/molbev/mst197>
- Telleria MT, Melo I, Dueñas M, Salcedo I, Beltrán-Tejera E, Rodríguez-Armas JL, Martín MP (2013a) *Sistotremastrum guttuliferum*: a new species from the Macaronesian islands. *Mycol Progress* 12:687–692
- Telleria MT, Melo I, Dueñas M, Larsson KH, Paz Martín MP (2013b) Molecular analyses confirm *Brevicellium* in Trechisporales. *IMA Fungus* 4:21–28
- Telleria MT, Dueñas M, Martín MP (2014) A new species of *Sistotremastrum* (Basidiomycota, Trechisporales) from Huinay Reserve. *Phytotaxa* 158:93–98
- Thiers B (2019) Index Herbariorum: a global directory of public herbaria and associated stuff [continuously updated]. New York Botanical Garden's Virtual Herbarium. <http://sweetgum.nybg.org/ih>
- Vilgalys R, Hester M (1990) Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several species of *Cryptococcus*. *J Bacteriol* 172:4238–4246
- Wu F, Yuan Y, Zhao CL (2015) *Porpomyces submucidus* (Hydnodontaceae, Basidiomycota), a new species from tropical China based on morphological and molecular evidence. *Phytotaxa* 230:61–68

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.